



## Archaea from the gut microbiota of humans: Could be linked to chronic diseases?

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**1 Archaea from the gut microbiota of humans: Could be linked to chronic diseases?**

2  
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**37 Conflicts of interest**

38 The authors declare that they have no conflicts of interest.  
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46        **Abstract**

47        Archaea comprise a unique domain of organisms with distinct biochemical and  
48        genetic differences from bacteria. Methane-forming archaea, methanogens, constitute  
49        the predominant group of archaea in the human gut microbiota, with  
50        *Methanobrevibacter smithii* being the most prevalent. However, the effect of  
51        methanogenic archaea and their methane production on chronic disease remains  
52        controversial. As perturbation of the microbiota is a feature of chronic conditions, such  
53        as cardiovascular disease, neurodegenerative diseases and chronic kidney disease,  
54        assessing the influence of archaea could provide a new clue to mitigating adverse  
55        effects associated with dysbiosis. In this review, we will discuss the putative role of  
56        archaea in the gut microbiota in humans and the possible link to chronic diseases.

57

58        **Keywords:** Archaea, gut microbiota, methane, chronic kidney disease, cardiovascular  
59        disease.

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69        **Highlights**

- 70           • In humans, the predominant Archaea are methanogens in the gastrointestinal  
71           system;
- 72           • Archaea may reduce ROS and TMAO production and intestinal permeability;
- 73           • Methane can indirectly act in a mechanism that regulates the antioxidant  
74           response.

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## 92 **Introduction**

93           Archaea (previously termed Archaeobacteria) display biochemical and genetic  
94           differences from bacteria [1,2]. Some are extremophiles and can survive in volcanic

95 mud, hot springs, salt lakes, and highly alkaline or acidic waters, as well as in a range of  
96 temperate environments, including agricultural soil and plants. A small percentage of  
97 archaea are a component of the human gut microbiota, along with viruses, protozoa,  
98 bacteria and fungi [3].

99 In general, the human intestinal microbiome has a range of functions that impact  
100 human health, including vitamin synthesis, barrier protection, immune system  
101 modulation, digestion of nutrients and protection against pathobionts [4]. In some  
102 circumstances, however, opportunistic pathogens can enter the host as a consequence of  
103 dysbiosis and or injuries or breaches in host barriers [5]. While these processes and  
104 effects are well described for some bacteria, little is known about the diversity of  
105 archaea in the vertebrate gut and influencing factors.

106 In humans, the predominant Archaea are methanogens (hydrogenotrophic), which  
107 anaerobically reduce carbon dioxide (CO<sub>2</sub>) to methane gas (CH<sub>4</sub>) (**Fig 1**). The most  
108 common methanogen in the gut microbiota is *Methanobrevibacter smithii*, with a  
109 prevalence of around 95.7% [6]. Compared to healthy individuals, Archaea appear  
110 overrepresented in patients with inflammatory bowel disease, human periodontal  
111 disease [7], obesity, cancer [8] and diverticulosis [9]. Consequently, interest in the role  
112 of methanogenic archaea in human disease processes has increased [3]. However, there  
113 is as yet no evidence that they could ever act as pathogens, and there remains a limited  
114 study of the role of Archaea in chronic diseases, such as cardiovascular disease (CVD),  
115 diabetes and chronic kidney disease (CKD). Evaluating the prevalence and functions of  
116 methanogens in the gut microbiota may provide vital information to determine the  
117 composition of normative gut microbiota. It may also indicate strategies to mitigate  
118 dysbiosis, a common feature in the above diseases. For example, it is noted that  
119 methylotrophic methanogens reduce trimethylamine-N-oxide (TMAO) production (a

120 marker of CVD) by TMA (trimethylamine) depletion in the gut, as some methanogens  
121 use TMA to produce CH<sub>4</sub> [10].

122 In this review, we will discuss the role of Archaea in the gut microbiota in humans  
123 and whether targeting Archaea may reduce uremic toxins produced by the gut  
124 microbiota, such as TMAO in chronic diseases.

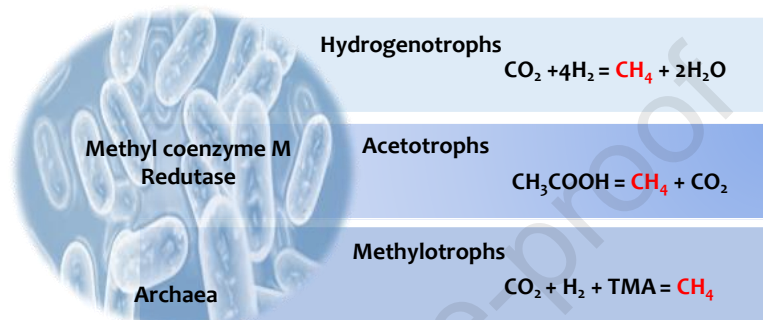
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### 126 ***Archaea structure and environment***

127 Archaea are unicellular, have no organelles and usually present DNA with a  
128 single circular chromosome. In addition to their similar size and shape, all these  
129 characteristics are identical to those found in bacteria [11]. However, archaea more  
130 closely resemble eukaryotes in DNA replication, transcription, damage repair and  
131 translation of genetic information [12]. Nonetheless, Archaea also possess some unique  
132 characteristics, such as the absence of peptidoglycans in the cell wall and membrane  
133 lipids formed by isoprenoid ether chains joined to sn-glycerol-1-phosphate, unlike  
134 bacteria, which have a membrane formed with chains of fatty acids linked to the  
135 glycerol-3-phosphate backbone via ester bonds [13].

136 Archaea are widely distributed in a range of habitats and may be extremophiles,  
137 thriving in extreme environments, such as hot springs, polar climates, acidic lakes,  
138 volcanoes, deserts, waste and water treatment plants and sites of highly concentrated  
139 environmental toxins. Archaeal extremophiles can phenotypically be divided into  
140 hyperthermophiles, inhabiting environments with extreme temperature conditions [2]  
141 piezophiles, flourishing in environments with high pressure; acidophiles and  
142 alkaliphiles, when required at low and high pH, respectively; and halophiles, growing  
143 optimally at high salt concentrations [14]. Archaea can also be divided into  
144 methanogens, which correspond to the phyla that inhabit anaerobic environments and

145 produce methane as a byproduct of metabolism [2]. Methanogens play a fundamental  
 146 role in the carbon cycle, being the only participating domain of methanogenesis. This  
 147 anaerobic respiration mechanism reduces carbon, oxygen, methyl or acetate with  
 148 hydrogen (as an electron donor) into methane through methyl-coenzyme M  
 149 reductase[15,16]. Methanogens can be divided into three main groups based on the  
 150 substrates used for methanogenesis (**Figure 1**).



151  
 152

153 **Figure 1. Methanogens are divided into three main physiological groups.** Hydrogenotrophs  
 154 are characterised by the absence of cytochromes and convert carbon dioxide and hydrogen into  
 155 methane using carbon as the terminal electron acceptor and dihydrogen as the leading electron  
 156 donor. Acetotrophs have cytochromes and use acetate as the only carbon source for methane  
 157 production. Methylootrophs also have cytochromes and can produce methane from hydrogen,  
 158 carbon dioxide, and methyl groups from compounds such as methanol and methylamines  
 159 (trimethylamine, dimethylamineylamine, monomethyl amine) and methyl-mercapto propionate,  
 160 among others.

161

162 Methanogens use substrates provided by syntrophic interactions (**Figure 2**) with

163 bacteria that carry out the anaerobic degradation of macronutrients, such as

164 carbohydrates, proteins and lipids [3]. For example, dietary carbohydrates undergo

165 hydrolysis by fermentative bacteria in the intestine to form pyruvate (final product of

166 glycolysis), which can be metabolised by secondary fermentative bacteria and produce

167 short-chain fatty acids (acetate, propionate and butyrate), organic acids (lactate, formate,

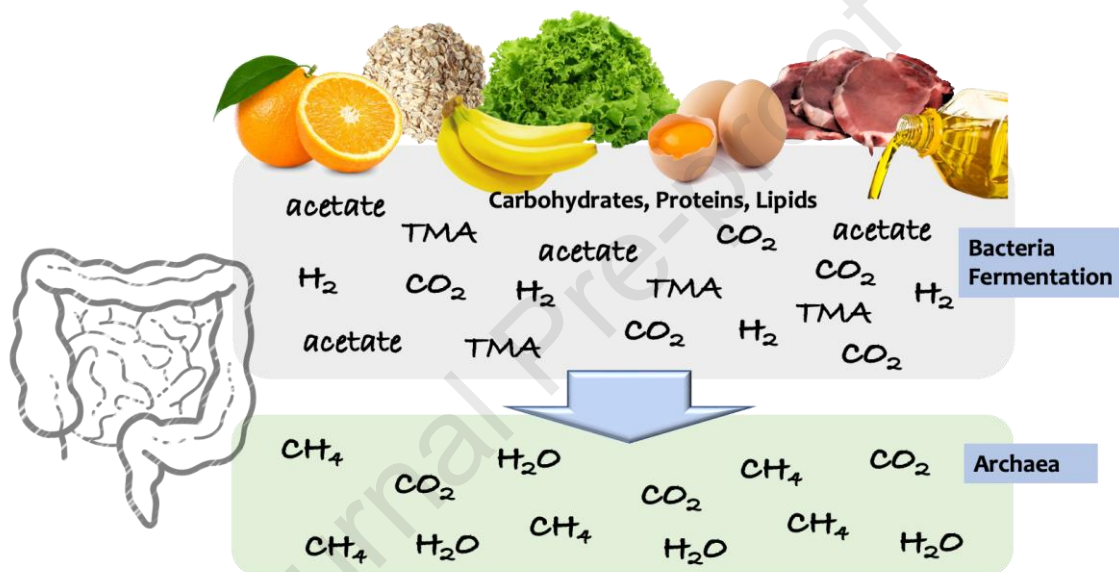
168 succinate), alcohols (ethanol) and gases (hydrogen and carbon dioxide) [17,18]. Such

169 products are used by hydrogenotrophs and acetotrophs or can later be decomposed and

170 converted by acetogenic bacteria [19].



171 Hydrogenotrophs can also utilise proteins as the byproduct of protein catabolism  
 172 that generates hydrogen [3]. The entire process of amino acid metabolism can generate  
 173 important substrates that fuel the growth of hydrogenotrophs, acetotrophs and  
 174 methylotrophs [3]. Finally, lipids contribute to methanogenesis through long-chain fatty  
 175 acids from lipid metabolism, which can be catabolised to short-chain fatty acids by  
 176 acetogenic bacteria, and glycerol, also from lipids, which can reach the glycolysis  
 177 pathway and contribute to hydrogenotrophs [3].



178  
 179 **Figure 2. Interactions between methanogens and bacteria in the gut microbiota.**  
 180 Fermentation of carbohydrates, proteins and lipids by bacteria results in the formation of  
 181 products such as carbon dioxide, hydrogen and acetate, which are metabolized by the archaea-  
 182 producing water (H<sub>2</sub>O), carbon dioxide (CO<sub>2</sub>), trimethylamine (TMA) and methane (CH<sub>4</sub>).  
 183

184 Archaea are a microorganism group that constitutes a relevant fraction of the  
 185 biomass present on our planet. They are a significant component of ocean microbiota  
 186 and are present in pelagic water samples, coastal surface waters[20,21], marine  
 187 sediment and terrestrial environments [22–24]. Archaea are involved in the biogenic  
 188 production of methane (CH<sub>4</sub>) and its oxidation, degradation of the protein [25], carbon  
 189 fixation [26,27], and sulfur, nitrogen and iron cycling [28] through several different

190 mechanisms using organic and inorganic compounds under aerobic and anaerobic  
191 conditions.

192 These methanogens are also related to anaerobic methane and short-chain carbon  
193 oxidation [29,30], which has already been proposed for bioremediation approaches [31].  
194 It is based on the capability of these anaerobic methane oxidizers (ANME) [30] to use  
195 the methane production pathway in a reverse direction [26] and thus degrade  
196 hydrocarbons [32–35]. This process is coupled to other processes, such as redox  
197 reactions, electron exchange and generation of energy realised by syntrophic bacteria  
198 [36]. It is evident that archaea play a vital environmental role and are critical factors in  
199 biogeochemical processes.

200 Archaeal methanogenesis is responsible for an important percentual of greenhouse  
201 gas generation since the enteric fermentation in ruminants produces methane [37,38].  
202 Indeed, the livestock sector (beef and dairy production) is responsible for 40% of  
203 greenhouse gas emissions [39], contributing significantly to global warming.

204 The level of released methane is supposed to increase rapidly due to climate  
205 warming and, consequently, the thawing of polar permafrost [40].

206 Methanogens are also directly involved in nitrogen fixation ( $N_2$ ), allowing most  
207 life forms to use a more suitable nitrogen compound from the atmosphere. Most archaea  
208 assimilate inorganic ( $NH_3$  and  $NO_3$ ) or organic compounds [41]. Urea seems to be an  
209 essential source of  $NH_3$  for ammonia-oxidizing thaumarchaeons in soil [42] and polar  
210 waters [43]. Another form of release of nitrogen gas in the atmosphere is from the  
211 denitrification process by oxidising inorganic compounds such as  $NO_2^-$  and  $NO_3^-$  [41].

212 Archaea (sulfidogens) belong to Euryarchaeota, and Crenarchaeota produce  
213 hydrogen sulfide ( $H_2S$ ) under anaerobic conditions, mainly in hydrothermal fields and

214 submarine vents [44]. Bacteria and archaea can reduce elementary sulfur to H<sub>2</sub>S and are  
215 widespread in the environment [45].

216 It is worth mentioning a hypothesis raised by Hallam et al. (2004), in which  
217 methane-producing relatives of Archaea would be able to completely reverse  
218 methanogenesis through the oxidation of methane to cellular carbon and energy. The  
219 anaerobic methane-oxidizing archaea would be responsible for this reversal [46]. The  
220 complete reversal of methane directly impacts the environment, playing an essential role  
221 in global warming and the inflow of greenhouse gases. Other researchers have also  
222 elucidated this hypothesis [47–49].

### 223 **Human archaeome: diversity, distribution and impacts in the gut**

224 Archaea represent approximately 1 to 2% of intestinal microorganisms in the  
225 human body and are mainly strictly anaerobic [50]; the most prevalent and abundant  
226 phyla are *Methanobacteriales* and *Methanomassiliicoccales* [9], with  
227 *Methanobrevibacter smithii* being the most found in the intestine reaching 99-100%  
228 among methane producers [51]. Regarding archaeological taxonomy, Rinke et al.  
229 (2021) recently proposed a standardised archaeal taxonomy from a phylogeny of 122  
230 proteins following the International Code of Prokaryote Nomenclature. Finally, the  
231 authors identified 16 archaeal phyla and reclassified three large monophyletic units  
232 from the ancient Euryarchaeota [52].

233 A recent study by La Cuesta-Zuluaga et al. (2021) has analysed 72 genomes of  
234 *Methanomassiliicoccales* and evaluated their presence in metagenomes derived from the  
235 human intestine, non-human intestine, and non-host environment. This research  
236 observed that archaea belonging to this order are generalists with a general preference  
237 for habitat, thus constituting their clade [53]. However, in humans, this group is not

238 joint, and when it is present, it is not found in high prevalence; in addition, diet can  
239 modulate its presence.

240 The methanogen group seems to be the only group involved in gut dysbiosis in  
241 humans, mainly *Methanobrevibacter smithii*, *Methanobrevibacter stadtmanae* and  
242 *Methanobrevibacter luminyensis* [54]. The methane produced by archaea (around  
243 0.35L/day) can be excreted by faeces ( $\approx 50\%$ ) or penetrate the intestinal mucosa and  
244 reach circulation. Methane production has been linked with the development of colon  
245 cancer [55,56] and diverticulosis [57,58], but this has not been supported by further  
246 research. However, strong evidence has indicated a relationship between methane and  
247 intestinal motility disorders [59–62].

248 Studies in dogs have shown that this gas acts directly on intestinal motility,  
249 delaying intestinal transit and thus contributing to constipation, probably via the  
250 cholinergic pathway (enteric nervous system) [63]. Pimentel et al. (2006) have further  
251 elucidated that the methane produced by archaea probably predisposes patients to the  
252 development of constipation by inciting segmental contractions (not proppant) [63].  
253 Kunkel et al. (2011) have corroborated these findings, and Rezaie et al. (2017) have  
254 found that methane can be used as a constipation marker [64,65].

255 However, other studies have attributed anti-inflammatory and cytoprotective  
256 effects to methane [66]. Boros et al. (2012) were pioneers in reporting the possible anti-  
257 inflammatory effect of methane in an ischemia-reperfusion study in inbred beagle dogs  
258 [67]. This study observed a significant reduction in reactive oxygen species (ROS)  
259 generation and modulation of leukocyte activation after methane administration [67].  
260 Mészáros et al. (2017) also observed similar results when analysing methane inhalation  
261 during ischemia and reperfusion in the small intestine of rats. Finally, they observed  
262 epithelial barrier preservation, reduction of permeability and improvement of local

263 microcirculation, leading to lower production of reactive oxygen and nitrogen species  
264 [68].

265 Methane can indirectly act in a mechanism that regulates the Keap1-Nrf2 system,  
266 from stimuli to the phosphatidylinositol 3-kinase (PI3K)-Akt pathway in macrophages  
267 stimulated by lipopolysaccharides [69]. It is crucial to notice that the Keap1-Nrf2  
268 system is the master endogenous defence mechanism with more than 250 syntheses of  
269 antioxidant enzymes. Wang et al. (2017)[70] have also reported that methane could act  
270 indirectly or directly in activating the Nrf2 pathway. It seems that byproducts of  
271 methane metabolism formed by electrophilic methyl groups can generate alterations and  
272 degrade Keap1, a protein in which Nrf2 remains coupled in the cytoplasm. Thus, the  
273 authors observed that methane might positively regulate transcriptional Nrf2 expression  
274 with a consequent increase in antioxidant enzyme expression [70]. Additionally,  
275 methane seems to increase the secretion of glucagon-like peptide 1 (GLP-1), a hormone  
276 responsible for increasing glucose-dependent pancreatic insulin secretion [71].

277 *Methanosphaera Stadtmanae* seems a potent immunostimulant, as *M. stadtmanae*  
278 binds to Toll-like receptor 8 (TLR8), triggers inflammasomes and exerts a more robust  
279 inflammatory response [72]. Furthermore, Archaea have been identified by their ability  
280 to activate dendritic cells derived from human monocytes, exerting strong inflammatory  
281 properties [72,73]. Studies on archaea and their role in inflammation and chronic  
282 diseases are still in their infancy, and humans' exact mechanism(s) is still not elucidated.

283

#### 284 **Could Archaea be a driving force for chronic diseases?**

285 The factors determining the possible relationship between methanogenic archaea  
286 and disease have remained undefined. Methanobrevibacter is not associated with  
287 diseases, and it is not clear its relationship with flatulence [74]. Evaluating high

288 methane emitters and low emitters, Kumpitsch et al. (2021) found that high emitters  
289 presented an increase in *Methanobrevibacter smithii*, which was associated with a high  
290 abundance of *Ruminococcaceae* and *Christensenellaceae*, which are involved with fiber  
291 degradation[75]. Supplementation with probiotics reduced the relative abundances of  
292 the *Methanobrevibacter*, which was related to lower production of gases [76].

293         Since a few years ago, researchers have observed a link between Archaea and  
294 neurological diseases, mainly multiple sclerosis (MS) [77–82]. Archaea may act in  
295 human genetics due to modulation of HLA gene expression that directly implies the  
296 RNA viroidal mRNA interference. Also, archaea may be involved with changes in the  
297 length of the non-coding region of genetic material, which performs an important role in  
298 the pathogenesis of autoimmune diseases such as systemic lupus erythematosus (SLE),  
299 rheumatoid arthritis (RA) and MS [77]. Another mechanism is related to the production  
300 of biofilm from the integration of nanoarchaea, prokaryotes, and viroids resulting in a  
301 new tissue phenotype in the neuronal and immune tissues, leading to these diseases[77].  
302 Archaea are also known for chronically activating the immune system and generating  
303 superantigens, which leads to the development of autoimmune diseases [77]. Whereas  
304 MS is an inflammatory disease mediated by the immune system with similar features to  
305 inflammatory bowel disease (IBD), the gut microbiota affects this disease as well as  
306 may affect MS and other neurological diseases [81] further. It is known there is an  
307 interaction between gut microbiota and the central nervous system through the  
308 microbiota-brain axis [79]. Related to this, some studies showed the interrelationship  
309 between MS and gut microbiota composition. In the patients with MS, there was a  
310 higher abundance of *Methanobrevibacteriaceae* (a genus in the Euryarchaeota) and a  
311 high prevalence of methane production from Archaea [78,80,82]. *Methanobrevibacter* is  
312 involved in the inflammatory process because of its capacity to recruit immune system

313 cells related to inflammation and its capacity to activate dendritic cells that are also  
314 involved in this process [78]. The same genus was positively correlated with TRAE 5, a  
315 regulator of cell T activation, which is overexpressed in MS [78].

316 Researchers have also considered a link between Archaea and body mass, where  
317 *Methanobrevibacter smithii* has been associated with both, obesity and malnutrition  
318 [83–85].

319 Methanogenic archaea are essential to remove excess H<sub>2</sub> from the human gut,  
320 reducing the efficiency of microbial fermentation and thereby the yield of energy. This  
321 prevents the buildup of H<sub>2</sub> and permits increased polysaccharide fermentation and short-  
322 chain fatty acids (SCFAs) production, which are well-known for their anti-inflammatory  
323 properties and modulation of the gut barrier[86] also, enhancing the availability of  
324 calories to the host, which is one of many mechanisms that link these bacteria with obesity  
325 [83,87]. In addition, bacterial NADH dehydrogenases are inhibited by the accumulation  
326 of H<sub>2</sub>, so the imbalance in the concentration of *M. Smithii* reflects, consequently, in the  
327 reduction of ATP production [88]. Studies have shown that obese patients have higher  
328 levels of methanogens and archaeal density in their faecal samples and methane breath  
329 tests [32,84,87,89–93]. This may be related to the production of SCFAs, improving  
330 fermentation efficiency and stimulating the lipogenesis process. Additionally, as the  
331 production of methane gas relates to constipation, this could take a long time for the  
332 intestine to absorb nutrients, resulting in weight gain[63,84,87,94–96]. However, the  
333 results are controversial. Armougom et al. (2009) found higher *M. smithii* in anorexic  
334 patients than in obese [83], and Wilder-Smith et al. (2019) found that obese patients had  
335 lower breath methane concentrations [97,98].

336 However, the results are controversial. Currently, studies demonstrate a link  
337 between gut microbiota and severe acute malnutrition, pointing to a dramatic depletion

338 of *M. smithii* [99,100]. Million et al. (2016) [99] demonstrated that *M. smithii* was not  
339 detected in children with severe acute malnutrition. This study proposed a theory of  
340 association between the oxidation of the intestinal environment and the depletion of  
341 anaerobic and methanogenic prokaryotes in malnutrition. Studies show that prokaryotic  
342 grow in an aerobic environment as long as the medium has adequate antioxidants [101].  
343 Furthermore, *M. smithii* can produce methane in an aerobic atmosphere using ascorbate,  
344 uric acid and glutathione [102]. Thus, these studies provided experimental evidence of a  
345 possible causal association between a lack of dietary antioxidants, anaerobic depletion  
346 (including *M. smithii*), and a severe reduction in substrate oxidation and energy uptake.

347 Subsequently, some studies [100,103] were carried out with the aim of clinically  
348 investigating the key role of *M. smithii* in severe acute malnutrition. Alou *et al.* (2017)  
349 [103] performed culturomics and metagenomics on stool samples from malnourished  
350 patients with kwashiorkor and healthy children and detected reduced diversity and  
351 depletion of *M. smithii*. The absence of this archaea was associated with the depletion of  
352 micronutrients (minerals, prebiotics, antioxidants) and the loss of a favourable  
353 environment for its growth.

354 In order to clarify whether the absence of *M. smithii* is the result of immaturity or  
355 a loss of microbiota. Camara, et al. (2021) [100] evaluated the presence of *M. smithii* in  
356 faeces collected from malnourished children at baseline and late to treatment. They  
357 observed that *M. smithii* was significantly associated with the absence of malnutrition and  
358 that intestinal dysbiosis presented during this comorbidity is not a result of immaturity  
359 but rather of a loss of this archaea.

360 Therefore, these studies provide speculation that the administration of potential  
361 probiotics associated with *M. smithii* may be an effective strategy for restoring the



362 intestinal microbiota in malnourished children [100,103]. Further studies are needed in  
363 order to more clearly elucidate this theory.

364 Archaea may also increase the risk of diabetes mellitus by impaired glucose  
365 tolerance and insulin resistance [87]. In a clinical trial, Cesario et al. (2014) found worse  
366 glycemic control with increased glycosylated haemoglobin (Hb1Ac) values in subjects who  
367 had methane production in their gut microbiota. Similar results can be observed in  
368 another study where methane-producing patients with diabetes mellitus type 1 had  
369 poorer glycemic control than nonmethane producers[84].

370 Several cardiovascular diseases (CVDs) and their risk factors have been identified  
371 as examples of interactions between the host and intestinal microbiota [104,105].

372 Studies suggest that the intestinal microbiome produces numerous metabolites that  
373 influence the host-microbiome composition and host health [106–108]. The most  
374 prominent molecule in this is trimethylamine N-oxide (TMAO) [109]. TMAO is formed  
375 from a bacterial precursor metabolite, trimethylamine (TMA). The latter reaches the  
376 bloodstream and undergoes metabolism in the liver, where it will be oxidised by hepatic  
377 flavine monooxygenases (more specifically FMO3) to TMAO, which has pro-  
378 atherogenic properties [109–111]. Indeed, rats receiving a diet rich in choline have  
379 shown high TMAO levels associated with leukocyte activation in endothelial cells,  
380 suggesting a role in the atherosclerotic process [112].

381 Plasma TMAO remains a potential biomarker for CVD development, even after  
382 controlling for traditional risk factors [109,113]. The increase in TMAO plasma levels  
383 is related to high proinflammatory cytokine levels, causing cardiac inflammation,  
384 fibrosis and atherosclerosis [35,114]. Elevated TMAO levels promote the activation of  
385 nuclear factor- $\kappa$ B (NF- $\kappa$ B), a transcription factor involved in activating an  
386 inflammatory pathway in human aortic endothelial and smooth muscle cells,

387 contributing to the atherosclerotic process. Additionally, TMAO significantly reduces  
388 the expression of Abcg5/8 (which acts to transport cholesterol out of the lumen) [115]  
389 and Niemann-Pick c1Like 1 (Npc1L1) (which is involved in cholesterol influx)  
390 [116,117]. Furthermore, it has been demonstrated that TMAO contributes to the  
391 inhibition of reverse cholesterol transport (RCT) [111] and the formation of foam cells  
392 and atherosclerotic plaques by increasing macrophage surface expression of scavenger  
393 receptor A (SRA) and the cluster of differentiation 36 (CD36) [106,118]. As broad-  
394 spectrum antibiotics suppress the pro-atherogenic effects, this underscores the  
395 importance of the intestinal microbiota in the metabolism of this toxin [111]. Strategies  
396 can be adopted to reduce TMAO, such as drugs to inhibit the FMO3 enzyme [119] or  
397 modulation directed to the bacterial domain.

398 *Candidatus (Ca.) Methanomethylophilus alvus* Mx1201 can produce methane  
399 using H<sub>2</sub> as an electron donor and TMA as an electron acceptor [120]. Another genus  
400 belonging to this family is *Methanomassiliicoccus luminyensis*, which has been shown  
401 to convert TMA to methane using H<sub>2</sub> *in vitro* [6]. However, although studies indicate  
402 that *M. luminyensis* is not a typical representative of the human intestine [121],  
403 members of Methanomassiliicoccales can use methylated amines, such as  
404 trimethylamine (TMA), as substrates for methanogenesis [122]. The observation that  
405 the Archaea domain can transform TMA into methane provides novel opportunities to  
406 reach promising strategies in the prevention/treatment of diseases [119], e.g., it has been  
407 suggested that methanogens may reduce the production of TMAO in patients with a  
408 hereditary defect in flavin-containing monooxygenase 3 [10]. Thus, the concept of  
409 Archaeobiotics arises, which would be applying specific archaeal intestinal strains  
410 capable of transforming TMA into methane and thus consequently reducing TMAO  
411 production, generating an inert gas for the host [123,124], being possible biotherapeutic

412 agents in the prevention and treatment of CVDs [53]. However, literature still includes  
413 few studies linking Archaea and CVD, and the presence of *Methanomassiliicoccales*  
414 that use TMA for growth is associated with lower faecal TMA concentrations [51].

415 Ramezani et al. (2018) experimentally tested the concept of archaeobiotics. To  
416 promote the persistence of archaea, antibiotics were initially administered to mice [125].  
417 Furthermore, mice were fed a diet rich in choline to increase TMAO levels, and after  
418 antibiotic therapy,  $10^8$  mesophilic methanogen cells were administered by single  
419 gavage. Surprisingly, *M. smithii* significantly reduced TMAO plasma levels compared  
420 to controls. Other methanogenic archaea (*Methanosarcina mazei*, *Methanomicrococcus*  
421 *blatticola*, *Methanohalophilus portucalensis*) reduced TMAO plasma levels. Still, this  
422 effect was detectable only in the first days of the experiment after gavage, probably due  
423 to low colonisation. After 30 days of the investigation, TMAO plasma levels were  
424 reduced, indicating a significant effect of *M. luminyensis* on TMA use. In addition to the  
425 direct application of archaea as live biotherapeutics, enzymes involved in  
426 methylotrophic metabolism could also be used. A well-known enzyme involved in this  
427 metabolism is trimethylamine methyltransferase (MttB), which transfers the methyl  
428 group from TMA to other methyltransferases. However, as this enzyme exerts its  
429 biological activity with the amino acid pyrrolysine (Pyl) in its active site [126], this  
430 would require a need to synthesise the amino acid pyrrolysine (one of the specificities  
431 of the genetics of TMA-consuming archaea).

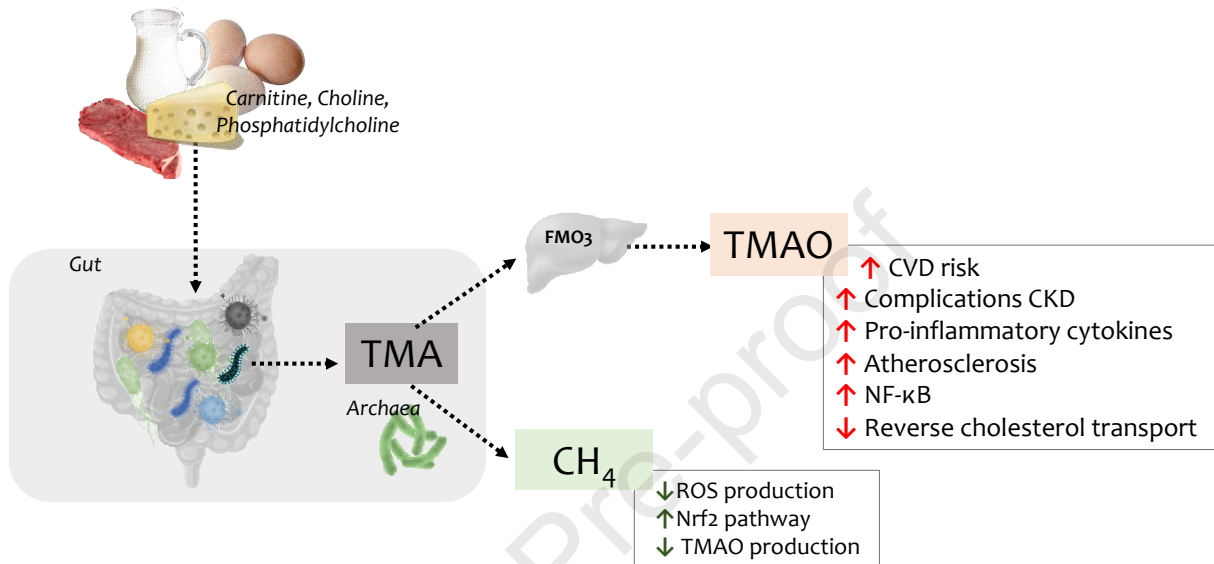
432 The salutogenic effects of archaea on the microbiome, metabolism, and host  
433 physiology need to be further studied *in vitro*, in animal models, and in humans.  
434 Specifically, archaea seem to confer beneficial effects in pro-inflammatory disease  
435 states (e.g., as observed with an accumulation of TMAO), such as trimethylaminuria

436 (TMAU), CVD and chronic kidney disease (CKD) and thus may have some potential as  
437 a therapeutic route to tackle inflammaging.

438 Although it is an important target due to the link with CVD, to the best of our  
439 knowledge, no studies have evaluated archaea in the gut of patients with CKD. Sumida  
440 et al. (2021) have hypothesised that microorganisms, including archaea, are present in  
441 the cell-free blood fraction of these patients. However, in a small study of 32  
442 hemodialysis patients, there was no evidence of intact archaeal DNA in the blood [127].  
443 Knobbe et al. (2020) evaluated kidney transplant recipients (KTx) to assess possible  
444 causes of dysbiosis and diarrhoea often encountered post-transplant [128]. The authors  
445 found a lower (28.6%) prevalence of *Methanobrevibacter smithii* in faeces from the  
446 KTx group than in faeces from a healthy control group (86.4%) [128]. Another  
447 important finding of this study was that the abundance of *Methanobrevibacter smithii*  
448 correlated positively with CH<sub>4</sub> concentrations in respiration and that CH<sub>4</sub>  
449 concentrations were lower in KTx [128]. A possible explanation for the reduction in  
450 *Methanobrevibacter smithii* in the faeces of KTx may be the more significant presence  
451 of sulfate-reducing bacteria [128]. These sulfate-reducing bacteria appear to compete  
452 for the use of H<sub>2</sub>, with *Methanobrevibacter smithii* becoming the predominant organism  
453 [129,130].

454 Despite this, it is already known that patients with CKD have dysbiosis with  
455 increased production of uremic toxins, including TMAO, which promotes oxidative  
456 stress, inflammation and CKD progression [131,132]. Recently, in an experimental  
457 model of adenine-induced CKD in rats, supplementation with  $\alpha$ -ketoacid for four weeks  
458 increased the abundance of Methanobrevibacter, which was negatively correlated with  
459 creatinine (Mo et al., 2021). Thus, patients with CKD accumulate TMAO in plasma  
460 [133–135], promoting CVD [136]. TMAO can then be absorbed by extrahepatic tissues

461 or excreted through urine, sweat, or breathing [111,137,138]. Indeed, TMAO is an  
 462 independent predictor of the burden of coronary atherosclerosis and mortality  
 463 independent of traditional risk factors related to CVD [139]. **Figure 3** shows the  
 464 possible role of archaea in humans.



465  
 466 **Figure 3.** Methylophilic methanogens can produce CH<sub>4</sub> from trimethylamine (TMA)  
 467 formed by fermentation of carnitine, choline, and phosphatidylcholine from the diet. On the other  
 468 hand, TMA can be absorbed and form trimethylamine-N-oxide (TMAO) in the liver. TMAO is  
 469 associated with cardiovascular disease (CVD) and chronic kidney disease (CKD) and increases  
 470 inflammation and atherosclerosis processes. Archaea reduce TMAO production. Methane (CH<sub>4</sub>)  
 471 produced by archaea may be of benefit.

472  
 473  
 474

### Conclusions and outlook

475 In recent decades, an understanding of the environmental role of archaea has increased  
 476 drastically. Starting with the simple idea that these microorganisms only lived in  
 477 extreme environments, it has now emerged that archaea are widespread even in  
 478 temperate habitats and participate in several critical processes and biogeochemical  
 479 cycles. Indeed, archaeal groups are vital for essential metabolism in nutrient cycles,  
 480 such as methanogenesis and methane oxidation. Recent studies have implied those  
 481 archaea may have a beneficial role by reducing ROS production, TMAO production and  
 482 intestinal permeability while activating the Nrf2 pathway [10,67,68]. It is possible that

483 targeting intestinal archaeal modulation could be a future strategy for managing  
484 dysbiosis and the high cardiovascular risk in patients with CKD. In the future, we  
485 believe new approaches in molecular microbial ecology will reveal even new and vital  
486 roles attributed to the archaeal domain.

487

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493

#### 494 **Conflicts of interest**

495 The authors declare that they have no conflicts of interest.

496

#### 497 **Contribution of authors**

498 All authors contributed significantly to the work's conception, literature collection and  
499 interpretation; participated in the writing or critical revision of the article in a manner  
500 sufficient to establish ownership of the intellectual content, and read and approved the  
501 version of the manuscript being submitted.

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#### 507 **References**

508

- 509 [1] C.R. Woese, G.E. Fox, Phylogenetic structure of the prokaryotic domain: the  
510 primary kingdoms, *Proc Natl Acad Sci U S A.* 74 (1977) 5088–5090.
- 511 [2] L. Eme, W.F. Doolittle, *Archaea*, 2015.
- 512 [3] K. Djemai, M. Drancourt, M.T. Alou, Bacteria and Methanogens in the Human  
513 Microbiome: a Review of Syntrophic Interactions, *Microbial Ecology.* 1 (2021)  
514 3. <https://doi.org/10.1007/s00248-021-01796-7>.
- 515 [4] A.L. Kau, P.P. Ahern, N.W. Griffin, A.L. Goodman, J.I. Gordon, Human  
516 nutrition, the gut microbiome and the immune system, *Nature.* 474 (2011) 327–  
517 336. <https://doi.org/10.1038/nature10213>.
- 518 [5] D. Ribet, P. Cossart, How bacterial pathogens colonize their hosts and invade  
519 deeper tissues, *Microbes and Infection.* 17 (2015) 173–183.  
520 <https://doi.org/10.1016/j.micinf.2015.01.004>.
- 521 [6] B. Dridi, M. Henry, A. El Khé chine, D. Raoult, M. Drancourt, High Prevalence  
522 of *Methanobrevibacter smithii* and *Methanosphaera stadtmanae* Detected in the  
523 Human Gut Using an Improved DNA Detection Protocol, *PLOS One.* 4 (2009).  
524 <https://doi.org/10.1371/journal.pone.0007063>.

- 525 [7] P.W. Lepp, M.M. Brinig, C.C. Ouverney, K. Palm, G.C. Armitage, D.A. Relman,  
526 Methanogenic Archaea and human periodontal disease, *Proc Natl Acad Sci U S*  
527 *A.* 101 (2004) 6176–6181. <https://doi.org/10.1073/PNAS.0308766101>.
- 528 [8] M. Cai, S. Kandalai, X. Tang, Q. Zheng, Contributions of Human-Associated  
529 Archaeal Metabolites to Tumor Microenvironment and Carcinogenesis,  
530 *Microbiol Spectr.* 10 (2022). <https://doi.org/10.1128/SPECTRUM.02367-21>.
- 531 [9] C.O. Guindo, M. 1 Drancourt, Grine G, Digestive tract methanodrome:  
532 physiological roles of human microbiota-associated methanogens, *Microbiology*  
533 *Pathogenesis.* (2020).
- 534 [10] J.F. Brugère, G. Borrel, N. Gaci, W. Tottey, P.W. O’Toole, C. Malpuech-  
535 Brugère, Archaeobiotics: Proposed therapeutic use of archaea to prevent  
536 trimethylaminuria and cardiovascular disease, *Gut Microbes.* 5 (2013).  
537 <https://doi.org/10.4161/gmic.26749>.
- 538 [11] C. Bräsen, D. Esser, B. Rauch, B. Siebers, Carbohydrate Metabolism in Archaea:  
539 Current Insights into Unusual Enzymes and Pathways and Their Regulation,  
540 *Microbiology and Molecular Biology Reviews.* 78 (2014) 89–175.  
541 <https://doi.org/10.1128/MMBR.00041-13>.
- 542 [12] E.P. Geiduschek, M. Ouhammouch, Archaeal transcription and its regulators,  
543 *Molecular Microbiology.* 56 (2005) 1397–1407. <https://doi.org/10.1111/j.1365-2958.2005.04627.x>.
- 544 [13] S.V. Albers, B.H. Meyer, The archaeal cell envelope, *Nature Reviews*  
545 *Microbiology.* 9 (2011) 414–426. <https://doi.org/10.1038/nrmicro2576>.
- 546 [14] P.H. Rampelotto, Extremophiles and extreme environments, *Life.* 3 (2013) 482–  
547 485. <https://doi.org/10.3390/life3030482>.
- 548 [15] Z. Lyu, N. Shao, A. Akinyemi, W. Whitman, Methanogenesis, *Current Biology.*  
549 28 (2018).
- 550 [16] S. Saengkerdsut, S.C. Ricke, Ecology and characteristics of methanogenic  
551 archaea in animals and humans, *Critical Reviews in Microbiology.* 40 (2014) 97–  
552 116. <https://doi.org/10.3109/1040841X.2013.763220>.
- 553 [17] M. Koch, J. Dolfing, K. Wuhrmann, A.J.B. Zehndert, Pathways of Propionate  
554 Degradation by Enriched Methanogenic Cultures, 1983.
- 555 [18] M. Tallefer, R. Sparling, Glycolysis as the Central Core of Fermentation, *Adv*  
556 *Biochem Eng Biotechnol.* 156 (2016) 55–78.  
557 [https://doi.org/10.1007/10\\_2015\\_5003](https://doi.org/10.1007/10_2015_5003).
- 558 [19] D. Ndeh, A. Rogowski, A. Cartmell, A.S. Luis, A. Baslé, J. Gray, I. Venditto, J.  
559 Briggs, X. Zhang, A. Labourel, N. Terrapon, F. Buffetto, S. Nepogodiev, Y.  
560 Xiao, R.A. Field, Y. Zhu, M.A. O’Neill, B.R. Urbanowicz, W.S. York, G.J.  
561 Davies, D.W. Abbott, M.C. Ralet, E.C. Martens, B. Henrissat, H.J. Gilbert,  
562 Complex pectin metabolism by gut bacteria reveals novel catalytic functions,  
563 *Nature.* 544 (2017) 65–70. <https://doi.org/10.1038/nature21725>.
- 564 [20] F. JA, M. K, D. AA, Novel major archaeobacterial group from marine plankton,  
565 *Nature.* 359 (1992) 167–169.
- 566 [21] E.F. Delong, Everything in moderation: Archaea as “non-extremophiles,” 1998.
- 567 [22] B.J. Baker, V. De Anda, K.W. Seitz, N. Dombrowski, A.E. Santoro, K.G. Lloyd,  
568 Diversity, ecology and evolution of Archaea, *Nature Microbiology.* 5 (2020)  
569 887–900. <https://doi.org/10.1038/s41564-020-0715-z>.
- 570 [23] K.G. Lloyd, L. Schreiber, D.G. Petersen, K.U. Kjeldsen, M.A. Lever, A.D. Steen,  
571 R. Stepanauskas, M. Richter, S. Kleindienst, S. Lenk, A. Schramm, B.B.  
572 Jorgensen, Predominant archaea in marine sediments degrade detrital proteins,  
573 *Nature.* 496 (2013) 215–218. <https://doi.org/10.1038/nature12033>.
- 574

- 575 [24] M.D. Dettling, J.B. Yavitt, H. Cadillo-Quiroz, C. Sun, S.H. Zinder, Soil-  
576 methanogen interactions in two peatlands (Bog, Fen) in central New York State,  
577 *Geomicrobiology Journal*. 24 (2007) 247–259.  
578 <https://doi.org/10.1080/01490450701456651>.
- 579 [25] A.L. Reysenbach, Y. Liu, A.B. Banta, T.J. Beveridge, J.D. Kirshtein, S.  
580 Schouten, M.K. Tivey, K.L. Von Damm, M.A. Voytek, A ubiquitous  
581 thermoacidophilic archaeon from deep-sea hydrothermal vents, *Nature*. 442  
582 (2006) 444–447. <https://doi.org/10.1038/nature04921>.
- 583 [26] S.J. Hallam, N. Putnam, C.M. Preston, J.C. Detter, D. Rokhsar, P.H. Richardson,  
584 E.F. DeLong, Reverse methanogenesis: Testing the hypothesis with  
585 environmental genomics, *Science* (1979). 305 (2004) 1457–1462.  
586 <https://doi.org/10.1126/science.11100025>.
- 587 [27] M. Könneke, D.M. Schubert, P.C. Brown, M. Hügler, S. Standfest, T.  
588 Schwander, L. Schada Von Borzyskowski, T.J. Erb, D.A. Stahl, I.A. Berg, D.M.  
589 Karl, Ammonia-oxidizing archaea use the most energy-efficient aerobic pathway  
590 for CO<sub>2</sub> fixation, *Proceedings of the National Academy of Sciences of*  
591 *the United States of America*. 111 (2014) 8239–8244.  
592 <https://doi.org/10.1073/pnas.1402028111>.
- 593 [28] P. Offre, A. Spang, C. Schleper, Archaea in biogeochemical cycles, *Annual*  
594 *Review of Microbiology*. 67 (2013) 437–457. <https://doi.org/10.1146/annurev-micro-092412-155614>.
- 596 [29] Y. Wang, G. Wegener, J. Hou, F. Wang, X. Xiao, Expanding anaerobic alkane  
597 metabolism in the domain of Archaea, *Nature Microbiology*. 4 (2019) 595–602.  
598 <https://doi.org/10.1038/s41564-019-0364-2>.
- 599 [30] V.J. Orphan, C.H. House, K.-U. Hinrichs, K.D. Mckeegan, E.F. Delong, Multiple  
600 archaeal groups mediate methane oxidation in anoxic cold seep sediments, *Proc*  
601 *Natl Acad Sci U S A*. 99 (2002) 7663–7668.
- 602 [31] M.J. Krzmarzick, D.K. Taylor, X. Fu, A.L. Mccutchan, Diversity and Niche of  
603 Archaea in Bioremediation, *Archaea*. (2018).  
604 <https://doi.org/10.1155/2018/3194108>.
- 605 [32] R.J. Basseri, B. Basseri, M. Pimentel, K. Chong, A. Youdim, K. Low, L. Hwang,  
606 E. Soffer, C. Chang, R. Mathur, Intestinal Methane Production in Obese  
607 Individuals Is Associated with a Higher Body Mass Index, 2012.
- 608 [33] R. Laso-Pérez, G. Wegener, K. Knittel, F. Widdel, K.J. Harding, V. Krukenberg,  
609 D. V. Meier, M. Richter, H.E. Tegetmeyer, D. Riedel, H.H. Richnow, L. Adrian,  
610 T. Reemtsma, O.J. Lechtenfeld, F. Musat, Thermophilic archaea activate butane  
611 via alkyl-coenzyme M formation, *Nature*. 539 (2016) 396–401.  
612 <https://doi.org/10.1038/nature20152>.
- 613 [34] G. Borrel, P.S. Adam, L.J. McKay, L.X. Chen, I.N. Sierra-García, C.M.K.  
614 Sieber, Q. Letourneur, A. Ghoulane, G.L. Andersen, W.J. Li, S.J. Hallam, G.  
615 Muyzer, V.M. de Oliveira, W.P. Inskeep, J.F. Banfield, S. Gribaldo, Wide  
616 diversity of methane and short-chain alkane metabolisms in uncultured archaea,  
617 *Nature Microbiology*. 4 (2019) 603–613. <https://doi.org/10.1038/s41564-019-0363-3>.
- 619 [35] S.C. Chen, N. Musat, O.J. Lechtenfeld, H. Paschke, M. Schmidt, N. Said, D.  
620 Popp, F. Calabrese, H. Stryhanyuk, U. Jaekel, Y.G. Zhu, S.B. Joye, H.H.  
621 Richnow, F. Widdel, F. Musat, Anaerobic oxidation of ethane by archaea from a  
622 marine hydrocarbon seep, *Nature*. 568 (2019) 108–111.  
623 <https://doi.org/10.1038/s41586-019-1063-0>.



- 624 [36] S. Scheller, H. Yu, G.L. Chadwick, S.E. McGlynn, V.J. Orphan, Artificial  
625 electron acceptors decouple archaeal methane oxidation from sulfate reduction,  
626 *Science* (1979). 351 (2016) 703–707. <https://doi.org/10.1126/science.aad7154>.
- 627 [37] J.G. Ferry, Fundamentals of methanogenic pathways that are key to the  
628 biomethanation of complex biomass, *Current Opinion in Biotechnology*. 22  
629 (2011) 351–357. <https://doi.org/10.1016/j.copbio.2011.04.011>.
- 630 [38] Y. Liu, W.B. Whitman, Metabolic, phylogenetic, and ecological diversity of the  
631 methanogenic archaea, in: *Ann N Y Acad Sci*, Blackwell Publishing Inc., 2008:  
632 pp. 171–189. <https://doi.org/10.1196/annals.1419.019>.
- 633 [39] FAO Statistical Yearbook 2021 - World Food and Agriculture - World |  
634 ReliefWeb, (n.d.). [https://reliefweb.int/report/world/fao-statistical-yearbook-](https://reliefweb.int/report/world/fao-statistical-yearbook-2021-world-food-and-agriculture)  
635 [2021-world-food-and-agriculture](https://reliefweb.int/report/world/fao-statistical-yearbook-2021-world-food-and-agriculture) (accessed January 26, 2022).
- 636 [40] R. MacKelprang, M.P. Waldrop, K.M. Deangelis, M.M. David, K.L. Chavarria,  
637 S.J. Blazewicz, E.M. Rubin, J.K. Jansson, Metagenomic analysis of a permafrost  
638 microbial community reveals a rapid response to thaw, *Nature*. 480 (2011) 368–  
639 371. <https://doi.org/10.1038/nature10576>.
- 640 [41] P. Cabello, M.D. Roldán, C. Moreno-Vivián, Nitrate reduction and the nitrogen  
641 cycle in archaea, *Microbiology (N Y)*. 150 (2004) 3527–3546.  
642 <https://doi.org/10.1099/mic.0.27303-0>.
- 643 [42] M. Tourna, M. Stieglmeier, A. Spang, M. Könneke, A. Schintlmeister, T. Urich,  
644 M. Engel, M. Schloter, M. Wagner, A. Richter, C. Schleper, *Nitrososphaera*  
645 *viennensis*, an ammonia oxidizing archaeon from soil, *Proc Natl Acad Sci U S A*.  
646 108 (2011). <https://doi.org/10.1073/pnas.1013488108>.
- 647 [43] L. Alonso-Sáez, A.S. Waller, D.R. Mende, K. Bakker, H. Farnelid, P.L. Yager,  
648 C. Lovejoy, J.-É. Tremblay, M. Potvin, F. Heinrich, M. Estrada, L. Riemann, C.  
649 Pedrós-Alió, S. Bertilsson, Role for urea in nitrification by polar marine Archaea,  
650 *Proc Natl Acad Sci U S A*. 109 (2012) 17989–17984.  
651 <https://doi.org/10.1073/pnas.1201914109>.
- 652 [44] A. Kletzin, Metabolism of Inorganic Sulfur Compounds in Archaea, *Archaea:*  
653 *Evolution, Physiology, and Molecular Biology*. (2007) 261–274.  
654 <https://doi.org/10.1002/9780470750865.ch23>.
- 655 [45] G. Muyzer, A.J.M. Stams, The ecology and biotechnology of sulphate-reducing  
656 bacteria, *Nature Reviews Microbiology*. 6 (2008) 441–454.  
657 <https://doi.org/10.1038/nrmicro1892>.
- 658 [46] S.J. Hallam, N. Putnam, C.M. Preston, J.C. Detter, D. Rokhsar, P.H. Richardson,  
659 E.F. DeLong, Reverse methanogenesis: Testing the hypothesis with  
660 environmental genomics, *Science* (1979). 305 (2004) 1457–1462.  
661 <https://doi.org/10.1126/science.1100025>.
- 662 [47] M. Cui, A. Ma, H. Qi, X. Zhuang, G. Zhuang, Anaerobic oxidation of methane:  
663 an “active” microbial process, *Microbiologyopen*. 4 (2015) 1–11.  
664 <https://doi.org/10.1002/MBO3.232>.
- 665 [48] P.H.A. Timmers, C.U. Welte, J.J. Koehorst, C.M. Plugge, M.S.M. Jetten, A.J.M.  
666 Stams, Reverse Methanogenesis and Respiration in Methanotrophic Archaea,  
667 *Archaea*. 2017 (2017). <https://doi.org/10.1155/2017/1654237>.
- 668 [49] Y. Wang, G. Wegener, J. Hou, F. Wang, X. Xiao, Expanding anaerobic alkane  
669 metabolism in the domain of Archaea, *Nature Microbiology*. 4 (2019) 595–602.  
670 <https://doi.org/10.1038/s41564-019-0364-2>.
- 671 [50] E. Garcia-Gutierrez, P.D. Cotter, Relevance of organ(s)-on-a-chip systems to the  
672 investigation of food-gut microbiota-host interactions, *Critical Reviews in*  
673 *Microbiology*. (2021) 1–26. <https://doi.org/10.1080/1040841X.2021.1979933>.

- 674 [51] G. Borrel, A. Mccann, J. Deane, M.C. Neto, D.B. Lynch, J.-F. Brugère, P.W.  
675 O'toole, Genomics and metagenomics of trimethylamine-utilizing Archaea in the  
676 human gut microbiome, *The ISME Journal*. 11 (2017) 2059–2074.  
677 <https://doi.org/10.1038/ismej.2017.72>.
- 678 [52] C. Rinke, M. Chuvochina, A.J. Mussig, P.A. Chaumeil, A.A. Davín, D.W. Waite,  
679 W.B. Whitman, D.H. Parks, P. Hugenholtz, A standardized archaeal taxonomy  
680 for the Genome Taxonomy Database, *Nat Microbiol*. 6 (2021) 946–959.  
681 <https://doi.org/10.1038/S41564-021-00918-8>.
- 682 [53] J. De La Cuesta-Zuluaga, T.D. Spector, N.D. Youngblut, R.E. Ley, Genomic  
683 Insights into Adaptations of Trimethylamine-Utilizing Methanogens to Diverse  
684 Habitats, Including the Human Gut Host-Microbe Biology, *MSystems*. 6 (2021).  
685 <https://doi.org/10.1128/mSystems.00939-20>.
- 686 [54] Y. Sereme, S. Mezouar, G. Grine, J.L. Mege, M. Drancourt, P. Corbeau, J. Vitte,  
687 Methanogenic Archaea: Emerging Partners in the Field of Allergic Diseases, *Clin  
688 Rev Allergy Immunol*. 57 (2016) 456–466. <https://doi.org/10.1007/s12016-019-08766-5>.
- 690 [55] A. Haines Jang Dilawari Geoffrey Metz Laurence Blendis, H. Wiggins Mrc, D.  
691 Epidemiology, M. Dunn Nutritional Laboratory Cambridge, Paper read at the 3rd  
692 international symposium on Detection and Prevention of Cancer, 1974.
- 693 [56] D. Polag, F. Keppler, Global methane emissions from the human body: Past,  
694 present and future, *Atmospheric Environment*. 214 (2019) 116823.  
695 <https://doi.org/10.1016/J.ATMOSENV.2019.116823>.
- 696 [57] S.-I. Jang, J.-H. Kim, Y.H. Youn, H. Park, S.I. Lee, J.L. Conklin, Relationship  
697 Between Intestinal Gas and the Development of Right Colonic Diverticula,  
698 *Journal of Neurogastroenterology and Motility*. 16 (2010) 418–423.  
699 <https://doi.org/10.5056/jnm.2010.16.4.418>.
- 700 [58] G.A. Weaver, J.A. Krause, T.L. Miller, M.J. Wolin, Incidence of methanogenic  
701 bacteria in a sigmoidoscopy population: an association of methanogenic bacteria  
702 and diverticulosis, *Gut*. 27 (1986) 698–704. <https://doi.org/10.1136/gut.27.6.698>.
- 703 [59] L. Hwang, A.E. Kimberly, L. Ae, R. Khoshini, A.E. Gil, M. Ae, A. Sahakian,  
704 A.E. Marc, M. Ae, V.P. Ae, M. Pimentel, Evaluating Breath Methane as a  
705 Diagnostic Test for Constipation-Predominant IBS, *Dig Dis Sci*. 55 (2010) 398–  
706 403. <https://doi.org/10.1007/s10620-009-0778-4>.
- 707 [60] G. Kim, F. Deepinder, W. Morales, L. Hwang, S. Weitsman, C. Chang, R.  
708 Gunsalus, M. Pimentel, *Methanobrevibacter smithii* Is the Predominant  
709 Methanogen in Patients with Constipation-Predominant IBS and Methane on  
710 Breath, *Dig Dis Sci*. 57 (2012) 3213–3218. <https://doi.org/10.1007/s10620-012-2197-1>.
- 712 [61] A.B. Sahakian, S.-R. Jee, M. Pimentel, Methane and the Gastrointestinal Tract,  
713 *Dig Dis Sci*. 55 (2010) 2135–2143. <https://doi.org/10.1007/s10620-009-1012-0>.
- 714 [62] K. Triantafyllou, C. Chang, M. Pimentel, Methanogens, methane and  
715 gastrointestinal motility, *Journal of Neurogastroenterology and Motility*. 20  
716 (2014) 31–40. <https://doi.org/10.5056/jnm.2014.20.1.31>.
- 717 [63] M. Pimentel, H.C. Lin, P. Enayati, B. van den Burg, H.-R. Lee, J.H. Chen, S.  
718 Park, Y. Kong, J. Conklin, Methane, a gas produced by enteric bacteria, slows  
719 intestinal transit and augments small intestinal contractile activity, *Am J Physiol  
720 Gastrointest Liver Physiol*. 290 (2006) 1089–1095.  
721 <https://doi.org/10.1152/ajpgi.00574.2004>.
- 722 [64] D. Kunkel, R.J. Basseri, M.D. Makhani, K. Chong, C. Chang, M. Pimentel,  
723 Methane on Breath Testing Is Associated with Constipation: A Systematic

- 724 Review and Meta-analysis, *Dig Dis Sci.* 56 (2011) 1612–1618.  
725 <https://doi.org/10.1007/s10620-011-1590-5>.
- 726 [65] A. Rezaie, M. Buresi, A. Lembo, H. Lin, R. McCallum, S. Rao, M. Schmulson,  
727 M. Valdovinos, S. Zakko, M. Pimentel, Hydrogen and Methane-Based Breath  
728 Testing in Gastrointestinal Disorders: The North American Consensus, *Nature*  
729 *Publishing Group.* 112 (2017). <https://doi.org/10.1038/ajg.2017.46>.
- 730 [66] S. Martinotti, F. Widdel, M. Boros, F. Keppler, Methane Production and  
731 Bioactivity-A Link to Oxido-Reductive Stress, *Front. Physiol.* 10 (2019) 1244.  
732 <https://doi.org/10.3389/fphys.2019.01244>.
- 733 [67] M. Boros, M. Ghyczy, D. Irces, G. Varga, T. Tokés, K. Kupai, C. Torday, J.  
734 Kaszaki, The anti-inflammatory effects of methane, *Critical Care Medicine.* 40  
735 (2012) 1269–1278. <https://doi.org/10.1097/CCM.0b013e31823dae05>.
- 736 [68] A.T. Mészáros, T. Büki, B. Fazekas, E. Tuboly, K. Horváth, M.Z. Poles, S.  
737 Szűcs, G. Varga, J. Kaszaki, M. Boros, Inhalation of methane preserves the  
738 epithelial barrier during ischemia and reperfusion in the rat small intestine,  
739 *Surgery (United States).* 161 (2017) 1696–1709.  
740 <https://doi.org/10.1016/j.surg.2016.12.040>.
- 741 [69] X. Zhang, N. Li, H. Shao, Y. Meng, L. Wang, Q. Wu, Y. Yao, J. Li, J. Bian, Y.  
742 Zhang, X. Deng, Methane limit LPS-induced NF- $\kappa$ B/ MAPKs signal in  
743 macrophages and suppress immune response in mice by enhancing  
744 PI3K/AKT/GSK-3 $\beta$ -mediated IL-10 expression OPEN, *Nature Publishing*  
745 *Group.* (2016). <https://doi.org/10.1038/srep29359>.
- 746 [70] D.X. Wang L, Yao Y, He R, Meng Y, Li N, Zhang D, Xu J, Chen O, Cui J, Bian  
747 J, Zhang Y, Chen G, Methane ameliorates spinal cord ischemia-reperfusion  
748 injury in rats: Antioxidant, anti-inflammatory and anti-apoptotic activity  
749 mediated by Nrf2 activation, *Free Radic Biol Med.* 103 (2017) 69–86.  
750 <https://doi.org/10.1016/J.FREERADBIOMED.2016.12.014>.
- 751 [71] R. Laverdure, A. Mezouari, M.A. Carson, N. Basiliko, J. Gagnon, A role for  
752 methanogens and methane in the regulation of GLP-1, *Endocrinology, Diabetes*  
753 *& Metabolism.* 1 (2018) e00006. <https://doi.org/10.1002/edm2.6>.
- 754 [72] X. Rao, H. Zhao, T. Vierbuchen, C. Bang, H. Rosigkeit, R.A. Schmitz, H. Heine,  
755 The human-associated archaeon *Methanosphaera stadtmanae* is recognized  
756 through its rna and induces Tlr8-Dependent nlrP3 inflammasome activation,  
757 *Frontiers in Immunology.* 8 (2017). <https://doi.org/10.3389/fimmu.2017.01535>.
- 758 [73] C. Bang, K. Weidenbach, T. Gutschmann, H. Heine, R.A. Schmitz, The intestinal  
759 archaea *Methanosphaera stadtmanae* and *Methanobrevibacter smithii* activate  
760 human dendritic cells, *PLoS ONE.* 9 (2014).  
761 <https://doi.org/10.1371/journal.pone.0099411>.
- 762 [74] J.A.A. van de Pol, N. van Best, C.A. Mbakwa, C. Thijs, P.H. Savelkoul, I.C. Ilja,  
763 M.W. Hornef, M. Mommers, J. Penders, Gut colonization by methanogenic  
764 archaea is associated with organic dairy consumption in children, *Frontiers in*  
765 *Microbiology.* 8 (2017). <https://doi.org/10.3389/fmicb.2017.00355>.
- 766 [75] C. Kumpitsch, F.P.S. Fischmeister, A. Mahnert, S. Lackner, M. Wilding, C.  
767 Sturm, A. Springer, T. Madl, S. Holasek, C. Högenauer, I.A. Berg, V. Schoepf,  
768 C. Moissl-Eichinger, Reduced B12 uptake and increased gastrointestinal formate  
769 are associated with archaeome-mediated breath methane emission in humans,  
770 *Microbiome.* 9 (2021). <https://doi.org/10.1186/S40168-021-01130-W>.
- 771 [76] M. Seo, J. Heo, J. Yoon, S.Y. Kim, Y.M. Kang, J. Yu, S. Cho, H. Kim,  
772 *Methanobrevibacter* attenuation via probiotic intervention reduces flatulence in

- 773 adult human: A non-randomised paired-design clinical trial of efficacy, PLoS  
774 ONE. 12 (2017). <https://doi.org/10.1371/JOURNAL.PONE.0184547>.
- 775 [77] R. Kurup, P.A. Kurup, A cholesterol and actinide dependent shadow biosphere of  
776 archaea and viroids in autoimmune diseases, Immunobiology. 217 (2012) 316–  
777 320. <https://doi.org/10.1016/J.IMBIO.2011.10.005>.
- 778 [78] S. Jangi, R. Gandhi, L.M. Cox, N. Li, F. von Glehn, R. Yan, B. Patel, M.A.  
779 Mazzola, S. Liu, B.L. Glanz, S. Cook, S. Tankou, F. Stuart, K. Melo, P. Nejad,  
780 K. Smith, B.D. Topçuoğlu, J. Holden, P. Kivisäkk, T. Chitnis, P.L. de Jager, F.J.  
781 Quintana, G.K. Gerber, L. Bry, H.L. Weiner, Alterations of the human gut  
782 microbiome in multiple sclerosis, Nat Commun. 7 (2016).  
783 <https://doi.org/10.1038/NCOMMS12015>.
- 784 [79] H. Tremlett, K.C. Bauer, S. Appel-Cresswell, B.B. Finlay, E. Waubant, The gut  
785 microbiome in human neurological disease: A review, Ann Neurol. 81 (2017)  
786 369–382. <https://doi.org/10.1002/ANA.24901>.
- 787 [80] A.I. Mirza, F. Zhu, N. Knox, J.D. Forbes, G. van Domselaar, C.N. Bernstein, M.  
788 Graham, R.A. Marrie, J. Hart, E.A. Yeh, D.L. Arnold, A. Bar-Or, J. O'Mahony,  
789 Y. Zhao, W. Hsiao, B. Banwell, E. Waubant, H. Tremlett, Metagenomic Analysis  
790 of the Pediatric-Onset Multiple Sclerosis Gut Microbiome, Neurology. 98 (2022)  
791 E1050–E1063. <https://doi.org/10.1212/WNL.0000000000013245>.
- 792 [81] J.D. Forbes, C.N. Bernstein, H. Tremlett, G. van Domselaar, N.C. Knox, A  
793 Fungal World: Could the Gut Mycobiome Be Involved in Neurological Disease?,  
794 Frontiers in Microbiology. 9 (2018) 3249.  
795 <https://doi.org/10.3389/FMICB.2018.03249>.
- 796 [82] J. Ochoa-Repáraz, L.H. Kasper, The influence of gut derived CD39 regulatory T  
797 cells in CNS demyelinating disease, Transl Res. 179 (2017) 126.  
798 <https://doi.org/10.1016/J.TRSL.2016.07.016>.
- 799 [83] F. Armougom, M. Henry, B. Vialettes, D. Raccach, D. Raoult, Monitoring  
800 Bacterial Community of Human Gut Microbiota Reveals an Increase in  
801 Lactobacillus in Obese Patients and Methanogens in Anorexic Patients, PLOS  
802 One. 4 (2009). <https://doi.org/10.1371/journal.pone.0007125>.
- 803 [84] R. Mathur, M. Amichai, K.S. Chua, J. Mirocha, G.M. Barlow, M. Pimentel,  
804 Methane and Hydrogen Positivity on Breath Test Is Associated With Greater  
805 Body Mass Index and Body Fat, J Clin Endocrinol Metab. 98 (2013).  
806 <https://doi.org/10.1210/jc.2012-3144>.
- 807 [85] M. Million, M. Maraninchi, M. Henry, F. Armougom, H. Richet, P. Carrieri, R.  
808 Valero, D. Raccach, B. Vialettes, D. Raoult, Obesity-associated gut microbiota is  
809 enriched in Lactobacillus reuteri and depleted in Bifidobacterium animalis and  
810 Methanobrevibacter smithii, International Journal of Obesity. 36 (2012) 817–825.  
811 <https://doi.org/10.1038/ijo.2011.153>.
- 812 [86] M. Esgalhado, J.A. Kemp, N.R.T. Damasceno, D. Fouque, D. Mafra, Short-chain  
813 fatty acids: a link between prebiotics and microbiota in chronic kidney disease,  
814 Future Microbiol. 12 (2017) 1413–1425. <https://doi.org/10.2217/FMB-2017-0059>.
- 815  
816 [87] G.M. Barlow, A. Yu, R. Mathur, Role of the gut microbiome in obesity and  
817 diabetes mellitus, Nutrition in Clinical Practice. 30 (2015) 787–797.  
818 <https://doi.org/10.1177/0884533615609896>.
- 819 [88] B.S. Samuel, A. Shaito, T. Motoike, F.E. Rey, F. Backhed, J.K. Manchester, R.E.  
820 Hammer, S.C. Williams, J. Crowley, M. Yanagisawa, J.I. Gordon, Effects of the  
821 gut microbiota on host adiposity are modulated by the short-chain fatty-acid

- 822 binding G protein-coupled receptor, Gpr41, Proc Natl Acad Sci U S A. 105  
823 (2008) 16767–16772. <https://doi.org/10.1073/PNAS.0808567105>.
- 824 [89] N.M. Delzenne, P.D. Cani, A. Everard, A.M. Neyrinck, L.B. Bindels, Gut  
825 microorganisms as promising targets for the management of type 2 diabetes,  
826 *Diabetologia*. 58 (2015) 2206–2217. <https://doi.org/10.1007/s00125-015-3712-7>.
- 827 [90] R.E. Ley, F. Bä Ckhed, P. Turnbaugh, C.A. Lozupone, R.D. Knight, J.I. Gordon,  
828 Obesity alters gut microbial ecology, Proc Natl Acad Sci U S A. 102 (2005)  
829 11070–11075.
- 830 [91] R. Mathur, K.S. Chua, M. Mamelak, W. Morales, G.M. Barlow, R. Thomas, D.  
831 Stefanovski, S. Weitsman, Z. Marsh, R.N. Bergman, M. Pimentel, Metabolic  
832 Effects of Eradicating Breath Methane Using Antibiotics in Prediabetic Subjects  
833 with Obesity, *Obesity*. 24 (2016) 576–582. <https://doi.org/10.1002/oby.21385>.
- 834 [92] D.P. Patil, D.P. Dhotre, S.G. Chavan, A. Sultan, D.S. Jain, V.B. Lanjekar, J.  
835 Gangawani, P.S. Shah, J.S. Todkar, S. Shah, D.R. Ranade, M.S. Patole, Y.S.  
836 Shouche, Molecular analysis of gut microbiota in obesity among Indian  
837 individuals, *J Biosci*. 37 (2012) 647–657. [https://doi.org/10.1007/s12038-012-](https://doi.org/10.1007/s12038-012-9244-0)  
838 9244-0.
- 839 [93] P.J. Turnbaugh, R.E. Ley, M.A. Mahowald, V. Magrini, E.R. Mardis, J.I.  
840 Gordon, An obesity-associated gut microbiome with increased capacity for  
841 energy harvest, *Nature*. 444 (2006) 1027–1031.  
842 <https://doi.org/10.1038/nature05414>.
- 843 [94] J. Jahng, I.S. Jung, E.J. Choi, J.L. Conklin, H. Park, The effects of methane and  
844 hydrogen gases produced by enteric bacteria on ileal motility and colonic transit  
845 time, *Neurogastroenterology and Motility*. 24 (2012).  
846 <https://doi.org/10.1111/j.1365-2982.2011.01819.x>.
- 847 [95] O. Maya-Lucas, S. Murugesan, K. Nirmalkar, L.D. Alcaraz, C. Hoyo-Vadillo,  
848 M.L. Pizano-Zárate, J. García-Mena, The gut microbiome of Mexican children  
849 affected by obesity, *Anaerobe*. 55 (2019) 11–23.  
850 <https://doi.org/10.1016/j.anaerobe.2018.10.009>.
- 851 [96] B.S. Samuel, J.I. Gordon, A humanized gnotobiotic mouse model of host-  
852 archaeal-bacterial mutualism, Proc Natl Acad Sci U S A. 103 (2006) 10011–  
853 10016.
- 854 [97] M. Million, M. Tidjani Alou, S. Khelaifia, D. Bachar, J.-C. Lagier, N. Dione, S.  
855 Brah, P. Hugon, V. Lombard, F. Armougom, J. Fromonot, C. Robert, C.  
856 Michelle, A. Diallo, A. Fabre, R. Guieu, C. Sokhna, B. Henrissat, P. Parola, D.  
857 Raoult, Increased Gut Redox and Depletion of Anaerobic and Methanogenic  
858 Prokaryotes in Severe Acute Malnutrition OPEN, Nature Publishing Group.  
859 (2016). <https://doi.org/10.1038/srep26051>.
- 860 [98] C.H. Wilder-Smith, S.S. Olesen, A. Materna, A.M. Drewes, Breath methane  
861 concentrations and markers of obesity in patients with functional gastrointestinal  
862 disorders, *United European Gastroenterology Journal*. 6 (2018) 595–603.  
863 <https://doi.org/10.1177/2050640617744457>.
- 864 [99] M. Million, M. Tidjani Alou, S. Khelaifia, D. Bachar, J.-C. Lagier, N. Dione, S.  
865 Brah, P. Hugon, V. Lombard, F. Armougom, J. Fromonot, C. Robert, C.  
866 Michelle, A. Diallo, A. Fabre, R. Guieu, C. Sokhna, B. Henrissat, P. Parola, D.  
867 Raoult, Increased Gut Redox and Depletion of Anaerobic and Methanogenic  
868 Prokaryotes in Severe Acute Malnutrition OPEN, Nature Publishing Group.  
869 (2016). <https://doi.org/10.1038/srep26051>.
- 870 [100] A. Camara, S. Konate, M. Tidjani Alou, A. Kodio, A.H. Togo, S. Cortaredona, B.  
871 Henrissat, M.A. Thera, O.K. Doumbo, D. Raoult, M. Million, Clinical evidence

- 872 of the role of *Methanobrevibacter smithii* in severe acute malnutrition, *Sci Rep.*  
873 11 (2021) 5426. <https://doi.org/10.1038/S41598-021-84641-8>.
- 874 [101] B. la Scola, S. Khelaifia, J.C. Lagier, D. Raoult, Aerobic culture of anaerobic  
875 bacteria using antioxidants: a preliminary report, *Eur J Clin Microbiol Infect Dis.*  
876 33 (2014) 1781–1783. <https://doi.org/10.1007/S10096-014-2137-4>.
- 877 [102] S. Khelaifia, J.C. Lagier, V.D. Nkanga, E. Guilhot, M. Drancourt, D. Raoult,  
878 Aerobic culture of methanogenic archaea without an external source of hydrogen,  
879 *Eur J Clin Microbiol Infect Dis.* 35 (2016) 985–991.  
880 <https://doi.org/10.1007/S10096-016-2627-7>.
- 881 [103] M.T. Alou, M. Million, S.I. Traore, D. Mouelhi, S. Khelaifia, D. Bachar, A.  
882 Caputo, J. Delerce, S. Brah, D. Alhousseini, C. Sokhna, C. Robert, B.A. Diallo,  
883 A. Diallo, P. Parola, M. Golden, J.C. Lagier, D. Raoult, Gut Bacteria Missing in  
884 Severe Acute Malnutrition, Can We Identify Potential Probiotics by  
885 Culturomics?, *Front Microbiol.* 8 (2017).  
886 <https://doi.org/10.3389/FMICB.2017.00899>.
- 887 [104] J.L. Griffin, X. Wang, E. Stanley, Metabolomics, Metabonomics, and the Gut  
888 Microbiome Does Our Gut Microbiome Predict Cardiovascular Risk? A Review  
889 of the Evidence From Metabolomics Metabolomics, *Circ Cardiovasc Genet.* 8  
890 (2015) 187–191. <https://doi.org/10.1161/CIRCGENETICS.114.000219>.
- 891 [105] K. Griffiths, B.B. Aggarwal, R.B. Singh, H.S. Buttar, D. Wilson, F. De Meester,  
892 Food Antioxidants and Their Anti-Inflammatory Properties: A Potential Role in  
893 Cardiovascular Diseases and Cancer Prevention, *Diseases.* (2016).  
894 <https://doi.org/10.3390/diseases4030028>.
- 895 [106] Z. Wang, E. Klipfell, B.J. Bennett, R. Koeth, B.S. Levison, B. Dugar, A.E.  
896 Feldstein, E.B. Britt, X. Fu, Y.M. Chung, Y. Wu, P. Schauer, J.D. Smith, H.  
897 Allayee, W.H.W. Tang, J.A. Didonato, A.J. Lusis, S.L. Hazen, Gut flora  
898 metabolism of phosphatidylcholine promotes cardiovascular disease, *Nature.* 472  
899 (2011) 57–65. <https://doi.org/10.1038/nature09922>.
- 900 [107] H. Craven, D. McGuinness, S. Buchanan, N. Galbraith, D.H. McGuinness, B.  
901 Jones, E. Combet, D. Mafra, P. Bergman, A. Ellaway, P. Stenvinkel, U.Z. Ijaz,  
902 P.G. Shiels, Socioeconomic position links circulatory microbiota differences with  
903 biological age, *Sci Rep.* 11 (2021). <https://doi.org/10.1038/S41598-021-92042-0>.
- 904 [108] D. Mafra, N.A. Borges, B. Lindholm, P.G. Shiels, P. Evenepoel, P. Stenvinkel,  
905 Food as medicine: targeting the uraemic phenotype in chronic kidney disease,  
906 *Nat Rev Nephrol.* 17 (2021) 153–171. <https://doi.org/10.1038/S41581-020-00345-8>.
- 907  
908 [109] W.H.W. Tang, Z. Wang, K. Shrestha, A.G. Borowski, Y. Wu, R.W. Troughton,  
909 A.L. Klein, S.L. Hazen, Intestinal microbiota-dependent phosphatidylcholine  
910 metabolites, diastolic dysfunction, and adverse clinical outcomes in chronic  
911 systolic heart failure, *Journal of Cardiac Failure.* 21 (2015) 91–96.  
912 <https://doi.org/10.1016/j.cardfail.2014.11.006>.
- 913 [110] N. Gaci, G. Borrel, W. Tottey, P.W. O’Toole, J.F. Brugère, Archaea and the  
914 human gut: New beginning of an old story, *World Journal of Gastroenterology.*  
915 20 (2014) 16062–16078. <https://doi.org/10.3748/wjg.v20.i43.16062>.
- 916 [111] R.A. Koeth, Z. Wang, B.S. Levison, J.A. Buffa, E. Org, B.T. Sheehy, E.B. Britt,  
917 X. Fu, Y. Wu, L. Li, J.D. Smith, J.A. Didonato, J. Chen, H. Li, G.D. Wu, J.D.  
918 Lewis, M. Warrier, J.M. Brown, R.M. Krauss, W.H.W. Tang, F.D. Bushman,  
919 A.J. Lusis, S.L. Hazen, Intestinal microbiota metabolism of l-carnitine, a nutrient  
920 in red meat, promotes atherosclerosis, *Nature Medicine.* 19 (2013) 576–585.  
921 <https://doi.org/10.1038/nm.3145>.

- 922 [112] M.M. Seldin, Y. Meng, H. Qi, W. Zhu, Z. Wang, S.L. Hazen, A.J. Lusis, D.M.  
 923 Shih, Trimethylamine N-Oxide Promotes Vascular Inflammation Through  
 924 Signaling of Mitogen-Activated Protein Kinase and Nuclear Factor- $\kappa$ B, *J Am*  
 925 *Heart Assoc.* 5 (2016). <https://doi.org/10.1161/JAHA.115.002767>.
- 926 [113] H.S. Nam, Gut Microbiota and Ischemic Stroke: The Role of Trimethylamine N-  
 927 Oxide, *Journal of Stroke.* 21 (2019) 151–159.  
 928 <https://doi.org/10.5853/jos.2019.00472>.
- 929 [114] R.-H. Chou, C.-Y. Chen, I.-C. Chen, H.-L. Huang, Y.-W. Lu, C.-S. Kuo, C.-C.  
 930 Chang, -Hsun Huang, J.-W. Chen, & S.-J. Lin, Trimethylamine N-oxide,  
 931 Circulating endothelial progenitor Cells, and endothelial Function in patients  
 932 with stable Angina, *Scientific Reports.* 9 (2019). [https://doi.org/10.1038/s41598-](https://doi.org/10.1038/s41598-019-40638-y)  
 933 [019-40638-y](https://doi.org/10.1038/s41598-019-40638-y).
- 934 [115] S. Stender, R. Frikke-Schmidt, B.G. Nordestgaard, A. Tybjaerg-Hansen, The  
 935 ABCG5/8 cholesterol transporter and myocardial infarction versus gallstone  
 936 disease, *J Am Coll Cardiol.* 63 (2014) 2121–2128.  
 937 <https://doi.org/10.1016/j.jacc.2013.12.055>.
- 938 [116] M.H. Janeiro, M.J. Ramírez, F.I. Milagro, J.A. Martínez, M. Solas, Implication of  
 939 Trimethylamine N-Oxide (TMAO) in Disease: Potential Biomarker or New  
 940 Therapeutic Target, *Nutrients.* 10 (2018). <https://doi.org/10.3390/nu10101398>.
- 941 [117] L. Jia, J.L. Betters, L. Yu, Niemann-Pick C1-Like 1 (NPC1L1) protein in  
 942 intestinal and hepatic cholesterol transport, *Annual Review of Physiology.* 73  
 943 (2011) 239–259. <https://doi.org/10.1146/annurev-physiol-012110-142233>.
- 944 [118] G. Falony, S. Vieira-Silva, J. Raes, Microbiology Meets Big Data: The Case of  
 945 Gut Microbiota-Derived Trimethylamine, *Annual Review of Microbiology.* 69  
 946 (2015) 305–321. <https://doi.org/10.1146/annurev-micro-091014-104422>.
- 947 [119] K. Fadhlou, M.-E. Arnal, M. Martineau, P. Camponova, B. Ollivier, P.W. O, J.-  
 948 F. Brugère, J.-F. Brugère J-FrancoisBrugere, Archaea, specific genetic traits, and  
 949 development of improved bacterial live biotherapeutic products: another face of  
 950 next-generation probiotics, *Applied Microbiology and Biotechnology.* 104 (2020)  
 951 4705–4716. <https://doi.org/10.1007/s00253-020-10599-8>.
- 952 [120] G. Borrel, H.M.B. Harris, W. Tottey, A. Mihajlovski, N. Parisot, E. Peyretailade,  
 953 P. Peyret, S. Gribaldo, P.W. O’toole, J.-F. Brugère, Genome Sequence of  
 954 “Candidatus Methanomethylophilus alvus” Mx1201, a Methanogenic Archaeon  
 955 from the Human Gut Belonging to a Seventh Order of Methanogens, *J Bacteriol.*  
 956 194 (2012) 6944–6945. <https://doi.org/10.1128/JB.01867-12>.
- 957 [121] G. Borrel, H.M. B Harris, N. Parisot, N. Gaci, W. Tottey, A. Mihajlovski, J.  
 958 Deane, S. Gribaldo, O. Bardot, E. Peyretailade, P. Peyret, P.W. O, J.-F. Brugère,  
 959 C.G. Borrel, H. Hmb, B. J-f, Genome Sequence of “Candidatus  
 960 Methanomassiliicoccus intestinalis” Issoire-Mx1, a Third Thermoplasmatales-  
 961 Related Methanogenic Archaeon from Human Feces, *Genome Announcements.* 1  
 962 (2013). <https://doi.org/10.1128/genomeA.00453-13>.
- 963 [122] K. Lang, J. Schuldes, A. Klingl, A. Poehlein, R. Daniel, A. Brune, New Mode of  
 964 Energy Metabolism in the Seventh Order of Methanogens as Revealed by  
 965 Comparative Genome Analysis of “Candidatus Methanoplasma termitum,” *Appl*  
 966 *Environ Microbiol.* 81 (2015) 1338–1352. [https://doi.org/10.1128/AEM.03389-](https://doi.org/10.1128/AEM.03389-14)  
 967 [14](https://doi.org/10.1128/AEM.03389-14).
- 968 [123] C. Bang, T. Vierbuchen, T. Gutschmann, H. Heine, R.A. Schmitz, Immunogenic  
 969 properties of the human gut-associated archaeon *Methanomassiliicoccus*  
 970 *luminyensis* and its susceptibility to antimicrobial peptides, *PLOS One.* 12  
 971 (2017). <https://doi.org/10.1371/journal.pone.0185919>.

- 972 [124] J.-F. Brugère, G. Borrel, N. Gaci, W. Tottey, P.W. O'toole, C. Malpuech-  
973 Brugère, Gut Microbes Archaeobiotics Proposed therapeutic use of archaea to  
974 prevent trimethylaminuria and cardiovascular disease, *Archaeobiotics, Gut*  
975 *Microbes*. 5 (2014) 5–10. <https://doi.org/10.4161/gmic.26749>.
- 976 [125] A. Ramezani, T.D. Nolin, I.R. Barrows, M.G. Serrano, G.A. Buck, R.  
977 Regunathan-Shenk, R.E. West III, P.S. Latham, R. Amdur, D.S. Raj, Gut  
978 Colonization with Methanogenic Archaea Lowers Plasma Trimethylamine N-  
979 oxide Concentrations in Apolipoprotein e<sup>-/-</sup> Mice OPEN, *Scientific Reports*. 8  
980 (2018). <https://doi.org/10.1038/s41598-018-33018-5>.
- 981 [126] C.R. Woese, G.J. Olsen, M. Ibba, D. Söll, A New UAG-Encoded Residue in the  
982 Structure of a Methanogen Methyltransferase, 1995.
- 983 [127] K. Sumida, J.F. Pierre, Z. Han, T.S. Mims, P.K. Potukuchi, M. Yuzefpolskaya,  
984 P.C. Colombo, R.T. Demmer, S. Datta, C.P. Kovesdy, Circulating Microbial  
985 Signatures and Cardiovascular Death in Patients With ESRD, *Kidney*  
986 *International Reports*. (2021). <https://doi.org/10.1016/j.ekir.2021.07.023>.
- 987 [128] T.J. Knobbe, R.M. Douwes, D. Kremer, J.C. Swarte, M.F. Eisenga, A.W.  
988 Gomes-Neto, M. van Londen, F.T.M. Peters, H. Blokzijl, I.M. Nolte, W.H.  
989 Hendriks, H.J.M. Harmsen, S.J.L. Bakker, Altered gut microbial fermentation  
990 and colonization with methanobrevibacter smithii in renal transplant recipients,  
991 *Journal of Clinical Medicine*. 9 (2020). <https://doi.org/10.3390/jcm9020518>.
- 992 [129] S.U. Christl, G.R. Gibson, J.H. Cummings, S.U. Christi, Role of dietary sulphate  
993 in the regulation of methanogenesis in the human large intestine, *Gut*. 33 (1992)  
994 1234–1238.
- 995 [130] S.U. Christl, P.R. Murgatroyd, G.R. Gibson, J.H. Cummings, Production,  
996 metabolism, and excretion of hydrogen in the large intestine, *Gastroenterology*.  
997 102 (1992) 1269–1277. [https://doi.org/10.1016/0016-5085\(92\)90765-q](https://doi.org/10.1016/0016-5085(92)90765-q).
- 998 [131] N.A. Borges, A.F. Barros, L.S. Nakao, C.J. Dolenga, D. Fouque, D. Mafra,  
999 Protein-Bound Uremic Toxins from Gut Microbiota and Inflammatory Markers  
1000 in Chronic Kidney Disease, *Journal of Renal Nutrition*. 26 (2016) 396–400.  
1001 <https://doi.org/10.1053/j.jrn.2016.07.005>.
- 1002 [132] E. Castillo-Rodriguez, R. Fernandez-Prado, R. Esteras, M.V. Perez-Gomez, C.  
1003 Gracia-Iguacel, B. Fernandez-Fernandez, M. Kanbay, A. Tejedor, A. Lazaro, M.  
1004 Ruiz-Ortega, E. Gonzalez-Parra, A.B. Sanz, A. Ortiz, M.D. Sanchez-Niño,  
1005 Impact of altered intestinal microbiota on chronic kidney disease progression,  
1006 *Toxins (Basel)*. 10 (2018). <https://doi.org/10.3390/toxins10070300>.
- 1007 [133] T.W. Meyer, T.H. Hostetter, *Uremia*, 2007.
- 1008 [134] C. Missailidis, J. Hällqvist, A.R. Qureshi, P. Barany, O. Heimbürger, B.  
1009 Lindholm, P. Stenvinkel, P. Bergman, Serum trimethylamine-N-Oxide is strongly  
1010 related to renal function and predicts outcome in chronic kidney disease, *PLoS*  
1011 *ONE*. 11 (2016). <https://doi.org/10.1371/journal.pone.0141738>.
- 1012 [135] R. Vanholder, A. Pletinck, E. Schepers, G. Glorieux, Biochemical and Clinical  
1013 Impact of Organic Uremic Retention Solutes: A Comprehensive Update, *Toxins*  
1014 *(Basel)*. 10 (2018) 33. <https://doi.org/10.3390/toxins10010033>.
- 1015 [136] S. Ito, M. Yoshida, Protein-Bound Uremic Toxins: New Culprits of  
1016 Cardiovascular Events in Chronic Kidney Disease Patients, *Toxins (Basel)*. 6  
1017 (2014) 665–678. <https://doi.org/10.3390/toxins6020665>.
- 1018 [137] F. Bäckhed, Meat-metabolizing bacteria in atherosclerosis, *Nature Medicine*. 19  
1019 (2013) 533–534. <https://doi.org/10.1038/nm.3178>.
- 1020 [138] M.A. Bain, R. Faull, G. Fornasini, R.W. Milne, A.M. Evans, Accumulation of  
1021 trimethylamine and trimethylamine-N-oxide in end-stage renal disease patients



- 1022           undergoing haemodialysis, *Nephrol Dial Transplant*. 21 (2006) 1300–1304.  
1023           <https://doi.org/10.1093/ndt/gfk056>.  
1024   [139] J.R. Stubbs, J.A. House, A.J. Ocque, S. Zhang, C. Johnson, C. Kimber, K.  
1025           Schmidt, A. Gupta, J.B. Wetmore, T.D. Nolin, J.A. Spertus, A.S. Yu, Serum  
1026           Trimethylamine-N-Oxide is Elevated in CKD and Correlates with Coronary  
1027           Atherosclerosis Burden, *J Am Soc Nephrol*. 27 (2016) 305–313.  
1028           <https://doi.org/10.1681/ASN.2014111063>.  
1029  
1030

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## Highlights

- In humans, the predominant Archaea are methanogens in the gastrointestinal system;
- Archaea may reduce ROS and TMAO production and intestinal permeability;
- Methane can indirectly act in a mechanism that regulates the antioxidant response.

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**Declaration of interests**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

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