

BioMine: A database for metazoan biomineralization proteins

Biomineralization is the process by which living organisms construct hard skeletons creating complex structures that range from specialized tissues such as bone or teeth to ecosystems such as coral reefs. Biominerals are composed of both inorganic minerals and proteins, which give them extra hardness and special attributes. Biomineralization proteins are also known to be associated with multiple bone disorders and are therefore of biomedical importance. Herein we describe BioMine, a biomineralization centric protein database. Availability and implementation: BioMine can be accessed at <http://biomine.net>, SQL dump, FASTA files and source code are available for download as well.

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9 **Introduction**

10 Biom mineralization is a process in which minerals form inside or outside the cells of a
11 variety of organisms (Lowenstam & Weiner 1989; Simkiss & Wilbur 1989). In animals,
12 these minerals are primarily calcium carbonates and calcium phosphates (Knoll 2003).
13 The majority of biominerals formed in bones, shells, skeletons and spicules are composed
14 of mineral crystals, however all biominerals contain various amounts of other proteins
15 that give these minerals extraordinary properties. The cell orchestrates the mineral
16 formation process through the expression and translocation of proteins that nucleate the
17 crystals either intracellularly or extracellularly. More importantly, the cell has to inhibit
18 mineral formation and crystal growth in unwanted sites (Kawasaki et al. 2009; Marin et
19 al. 1996). Both nucleation and inhibition can be achieved through multiple cellular
20 mechanisms. For example, the cell will produce enzymes that modify proteins by
21 breaking them into smaller peptides (Qin et al. 2004) thus changing their function. The
22 cell is able to tightly regulate the biomineralization process by molecular modification
23 (e.g. adding sugars or other moieties) and regulation of ion transport across membranes
24 (Qin et al. 2004; Saavedra 1994; Sarashina et al. 2006). Such modifications to
25 biomineralization-associated proteins will determine how they interact with other
26 proteins, other cells, and with the biomineral in general. Biominerals are essential to the
27 survival of a broad range of animal taxa because they deliver protection against
28 predation, act as energy storage, provide support and unique optical properties (Addadi et
29 al. 2006). In particular, biomineralization plays a pivotal role in multiple human diseases
30 and other pathological phenomena such as coronary artery calcification (Atlan et al.
31 1997; Collette et al. 2010; Fisher et al. 2001; Lopez et al. 1992; Rousselle & Heymann
32 2002; Salih et al. 1996; Wallin et al. 2001; Westbroek & Marin 1998; Yang et al. 2002).
33 A growing interest in bio-inspired materials has generated a large body of work that uses
34 proteins and other biological scaffolds for *in vitro* mineralization and synthetic materials
35 (Chiu et al. 2012; Perry et al. 2009).
36
37 The process of biomineralization is ubiquitous throughout the animal tree. Such
38 distribution has generated speculation about the origin of metazoan biomineralization and

39 its evolutionary history. Biomineralization is a complex process that relies on multiple
40 cellular pathways (Knoll 2003; Marin et al. 1996). Many of the studied biomineralization
41 proteins are part of other important processes such as cell adhesion, extracellular matrix
42 organization and immune functions (Bryden et al. 1999; Clendenon et al. 2009). This
43 evidence favors the idea that biomineralization independently evolved in multiple phyla
44 using pre-existing pathways in the early eumetazoan ancestor. It can also be argued that
45 biomineralization was present in the early eumetazoan ancestor yet various parts of the
46 pathway were lost in several animal lineages. According to fossil evidence and when
47 mapped onto a phylogeny, carbonate skeletons seem to have evolved at least 20 different
48 times in metazoans (Knoll 2003). If biomineralization evolved multiple times, it is
49 relevant to understand which components of the process exactly underwent innovations.
50 Since biomineralization is an active process, it requires 1) targeted localization of
51 calcium and carbonate, 2) an organic matrix as a template for the mineral nucleation, 3)
52 growth, and 3) efficient inhibitors in order to stop undesired calcification or even
53 formation of the mineral (Jackson et al. 2010; Marin et al. 1996). When all these different
54 requirements are taken into account, it seems unlikely that such diverse biochemical
55 processes involved in metazoan biomineralization evolved independently more than 20
56 times. A process such as transport is quite conserved across animal lineages and it shows
57 a clear history of gene duplication events (Dean et al. 2001; Saier et al. 2009). Such
58 complexity presents us with a conundrum. While the biomineralization process, with
59 different minerals and methods of calcification and clear evolutionary novelties, is found
60 across multiple animal phyla (Jackson et al. 2007a; Jackson et al. 2006; Jackson et al.
61 2009; Jackson et al. 2007b; Marin et al. 2000; Marin & Luquet 2004; Marin et al. 2008;
62 Marin et al. 1996), it also remains that many parts of biomineralization pathways must be
63 conserved.

64

65 As a first step in tackling such questions in the evolution of animal biomineralization, we
66 have created a database to accumulate, annotate and curate biomineralization proteins and
67 protein-coding sequences. The database aims to serve the community by bridging the gap
68 between the few identified biomineralization proteins, and the unannotated plethora of
69 Expressed Sequence Tags (ESTs), draft genome gene models and next-generation

70 sequencing datasets. We employed various bioinformatics techniques using domain-
71 based searches to collect and identify novel biomineralization proteins in metazoans. We
72 hope that due to the increasing surge of sequence information along with broad
73 phylogenetic representation in the public domain, a clearer picture of the evolutionary
74 history of biomineralization proteins will emerge, rendering BioMine as a dynamic
75 platform to answer not only fundamental questions in animal evolution but also about the
76 process of biomineralization in particular lineages.

77

78 **Methods**

79 **Biomineralization proteins list**

80 We carried out a wide primary literature and database survey in order to compile a list of
81 proteins that are functionally implicated biomineralization in animals. Specifically, we
82 included data from scleractinian corals, calcareous sponges, gastropod and bivalve
83 molluscs, crustaceans, echinoderms, and vertebrates. Additional sequences were collected
84 from the AMIGO Gene Ontology database (Carbon et al. 2009). The complete
85 biomineralization gene list is accessible through the BioMine web application
86 (<http://biomine.net/>). These already annotated sequences were used as a seed to search for
87 related biomineralization proteins in undocumented taxa or new sequence databases, and
88 were stored in a pre-computed BLAST search database. After building the list of
89 candidate proteins, each protein could be traced back to an original publication where it
90 was described.

91

92 **Pfam domain search and protein homolog identification**

93 To further improve the search strategy, using the Pfam database, we scanned for
94 conserved protein domains in the proteins we gathered from primary literature (Finn et al.
95 2008). The identified domains in the already known biomineralization proteins were
96 scanned against 6-frame translations of ESTs and protein sequences from dbEST and nr
97 databases (NCBI) of the taxa Cnidaria, Mollusca, Echinodermata and Vertebrata using
98 the HMMER 3 package (Eddy 2008; Eddy 2011). The tool used for the translation was
99 sixpack from the EMBOSS package (Rice et al. 2000).

100

101 **BLAST Searches**

102 In addition to the domain searches, we conducted BLASTp searches for all the proteins in
103 the seed database against nr and against the 6-frame translations of the dbEST for the
104 selected taxa. For filtering the results, we only considered hits that match e-value >
105 0.000001 and bitscore > 150 to be significant.

106

107 **BioMine construction**

108 In order, to organize all the data in a searchable platform, we constructed a web
109 application that enables us to search the results and to submit new sequences into the
110 database. BioMine is written in PHP and Perl, and relies on MySQL for relational
111 information Source code for BioMine is under GPL v3 at
112 <http://code.google.com/p/biomine/>. The MySQL database contains the results of all the
113 HMMER results in addition to the BLAST results and FASTA files of all sequences can
114 be downloaded from the website.

115

116 **Results**

117 After assembling the initial list of biomineralization-related proteins, we identified
118 putative homologs of given candidate genes in calcifying lineages (molluscs, cnidarians,
119 arthropods and echinoderms). A protein domain search was initiated on our candidate list
120 of 472 proteins, based on Pfam models using HMMER 3 (Eddy 2008). In the Pfam
121 search 198 domain families were found to be linked to biomineralization. The search
122 results were stored in a relational database linking the detected domains with the species
123 in addition to detected orthologs for every particular protein.

124 Below we describe two potential scenarios for the use of BioMine by the scientific
125 community.

126 Use case 1:

- 127 1) A user prepares a list of proteins from a newly sequenced organism.
- 128 2) The user submits the protein list to BioMine through the web interface.
- 129 3) BioMine generates potential matching biomineralization proteins in the submitted
130 dataset, together with the publications in which these similar proteins have been
131 described. In addition, a predicted protein-protein interaction network will be
132 generated if the submitted protein list can be decomposed successfully to Pfam
133 domains. These domains are obtained from the Pfam database, which contains
134 curated conserved domains of various functions

135 Use case 2:

- 136 1) A user is already working with a known biomineralization protein and is doing
137 functional work, i.e. the user is doing whole mount in situ gene expression
138 research in a given organism and finds it hard to explain the observed expression
139 pattern. Thus, the user thinks there could be other proteins involved.
140 2) The user submits his protein (as sequence or as a gi identifier or uniprot id) to
141 BioMine.
142 3) The user gets back a list of potential interacting proteins and possible paralogs
143 restricted to a taxon of his choice if needed.
144

145 **Discussion and Conclusion**

146 By combining thorough literature scrutiny with similarity searches we were able to
147 construct a large dataset of biomineralization-related proteins. BioMine proved useful in
148 annotating sequence data from non-model organisms involved in the particular process of
149 biomineralization. The ability to always link back to the primary literature provides a
150 unique opportunity to the investigator to directly examine the experimental evidence that
151 deemed a particular protein as biomineralization-associated. We believe this should fast-
152 forward research in non-model systems by knowledge transfer from model species in
153 biomineralization research. By providing a BLAST interface and downloadable versions,
154 we are certain that biomineralization researchers can benefit from BioMine. BioMine is a
155 dedicated database containing manually curated biomineralization proteins from all
156 mineralizing animal taxa that is presented through a user-friendly fully searchable
157 interface.
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