

Semi-Classical Signal Analysis Method with Soft-Thresholding for MRS denoising

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Synopsis

A Semi-Classical Signal Analysis (SCSA) method with soft thresholding is proposed for MRS denoising. The SCSA takes advantage of the pulse-shaped MRS spectrum to decompose both real and imaginary parts, into localized basis given by squared eigenfunctions of the Schrödinger operator. An optimization-based soft-threshold is provided to find optimal semi-classical parameters, for both the real and imaginary parts of the MRS signal. The optimal SCSA parameters discard the eigenfunctions representing noise from the noisy spectrum, and conserve the eigenfunctions representing the useful information. The obtained in-vivo results show the efficiency of the SCSA with soft thresholding in removing noise and conserving metabolite signals.

Introduction

Most of the biomedical signals have a pulse-shaped form [1], which reflects biological or chemical activities happening within the human body. Magnetic Resonance Spectroscopy (MRS) signals are good examples of pulse-shaped signals that are widely used for medical diagnosis and monitoring during surgical operations. However, these signals are subject to noise contamination, due to many interferences, such as electronic equipment noise coming from the power line, patient motion, and others. Several MRS denoising methods have been proposed [2,3]. For instance, a recent work employed the Semi-Classical Signal Analysis (SCSA) to reduce the noise in MRS spectra. However, this algorithm deals only with the real part of the MRS spectrum. Moreover, the imaginary part of the MRS signal includes some undershoots (Negative peaks) which makes the basic SCSA unsuitable for denoising complex MRS. In this work, an extension of the SCSA algorithm is proposed to denoise both real and imaginary parts of the MRS signals, which uses a soft threshold to optimally reduce the noise while preserving the metabolites.

Material and Methods

1. The Dataset

Several single voxel spectroscopy in vivo data from frontal lobe region are collected from a healthy volunteer, using the following parameters: TE/TR=30/2000ms, 1024 points, 2500Hz bandwidth. The voxel size is set to 20 × 20 × 20 mm³ and the numbers of averages (Nex) are varied from 2 to 16. The SCSA method is applied to reduce the noise contribution prior to quantification of Choline (Cho), Creatine (Cr), Lactate (Lac), N-Acetyl-Aspartate (NAA) metabolite peaks and the calculations of NAA/Cr and Cho/Cr, and Lac/Cr ratios.

2. The SCSA Method

The noisy MRS $z(f)$ spectrum is first split into the real part $z_r(f)$ and imaginary part $z_i(f)$. Each of these two signals is considered as potential of a Schrödinger operator of the eigenvalue problem defined as follows:

$$-h_l^2 \frac{d^2 \psi(f)}{df^2} - z_l(f) \psi(f) = \mu \psi(f)$$

where $h_l > 0$, $l = \{r, i\}$ defines the real part $z_r(f)$ and the imaginary part $z_i(f)$ of the MRS signal respectively.

The denoised MRS signal $z_h(f)$ is given by:

$$z_h(f) = z_{r,h}(f) + i z_{i,h}(f)$$

such that :

$$z_{r,h}(f) = 4h_r \sum_{k=1}^{N_{r,h}} \sqrt{(-\mu_{r,kh})} \psi_{r,kh}^2(f) \quad \text{and} \quad z_{i,h}(f) = 4h_i \sum_{k=1}^{N_{i,h}} \sqrt{(-\mu_{i,kh})} \psi_{i,kh}^2(f)$$

where μ and $\psi(f)$ refer to the negative eigenvalues and associated L^2 -normalized eigenfunctions of the Schrödinger operator respectively.

3. The soft thresholding

The objective of this work is to find optimal values for the parameters h_i and h_r for efficient denoising. Therefore, an optimization-based thresholding is used with an adaptive selection of the optimization parameters depending on the shape of the signal and the number of peaks in the signal. This optimization is based on three steps:

1. Detect the location of the positive and negative peaks,
2. Split the signal into a set of smaller signals such that each has one or two peaks (see Figure 2),
3. Denoise each sub-signal alone by minimizing the cost function $J(h)$.

For instance, the cost function to find the optimal semi-classical parameter h_i and h_r for a specific split of the real or imaginary part is defined as follows :

$$J(h) = \begin{cases} \frac{STD\left(\frac{d^2 z_{l,h}(f_A)}{df^2}\right)}{STD\left(\frac{d^2 z_l(f_A)}{df^2}\right)} & , \quad \text{if } \rho < C \\ \infty & \text{elsewhere} \end{cases}$$

where $l = \{r, i\}$. $f_A \in A$, where A is an interval that describe the noisy zone. The STD refers to the Standard Deviation. C is the maximum threshold allowed for peak attenuation defined as:

$$\rho = \frac{|\max(z_{l,h}) - \max(z_l)|}{|\max(z_l(f))|}$$

Results and Discussion

The proposed SCSA with soft-thresholding has been tested on localized MRS in vivo data. As shown in the result with $Nex=4$ (Figure 3), the proposed SCSA efficiently reduces noise while preserving the metabolite peaks. Figure 3A shows the comparison of real parts of the original (blue) and the de-noised (red) spectra along with zoomed metabolite region shown in Figure 3C. Figure 3B displays the comparison of imaginary parts of the original (blue) and de-noised (red) spectra and zoomed metabolite region shown in Figure 3D.

The quantification results in terms of SNR, computed as the ratio of NAA peak area to the standard deviation of noise, and the metabolite ratios before and after de-noising are shown in given in Table 1. The AMARES [4] peak fitting method is used for the analysis of the NAA, Cr, Cho, and Lac peaks. The results show considerable improvement in the SNR and accurate metabolite ratios after denoising compared to the literature [5].

Conclusion

The new SCSA method with adaptive thresholding achieves efficient MRS signal denoising while preserving the metabolite peaks, demonstrated by the efficient noise reduction and metabolites preservation. The obtained results are very encouraging and show the potential of the SCSA with thresholding for general pulse-shaped signal denoising.

Acknowledgements

Research reported in this publication was supported by King Abdullah University of Science and Technology (KAUST).

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Figures

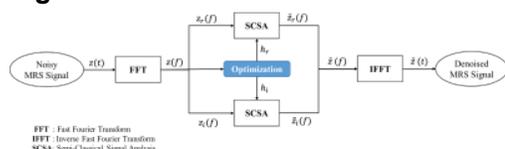


Figure 1: Complex MRS signal denoising using SCSA.

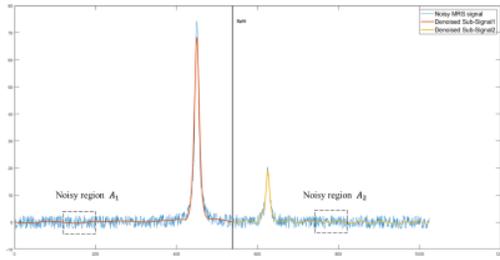


Figure 2: Example of signal splitting and denoising.

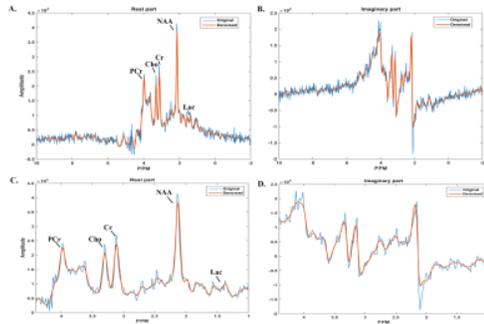


Figure 3: Result with in vivo data ($Nex = 4$) comparing real (A. and metabolite region zoomed in C.) and imaginary parts (B. and metabolite region zoomed in D.) of original (blue) with the de-noised spectra (red).

	SNR		NAA/Cr		Cho/Cr		Lac/Cr	
	Before	After	Before	After	Before	After	Before	After
$Nex=2$	7.1	14.9	1.55	1.78	0.92	0.89	0.21	0.18
$Nex=4$	11.3	24.3	1.53	1.62	0.88	0.87	0.21	0.24
$Nex=8$	15.5	55.8	1.61	1.61	0.86	0.85	0.22	0.22
$Nex=16$	20.3	65.5	1.61	1.61	0.86	0.86	0.22	0.22

Table 1: Invivo results showing the SNR values and metabolites ratios before and after SCSA denoising.