

## LETTERS

# Large Number of Ultraconserved Elements Were Already Present in the Jawed Vertebrate Ancestor

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Stephen (2008) identified 13,736 ultraconserved elements (UCEs) in placental mammals and investigated their evolution in opossum, chicken, frog, and fugu. They found that there was a massive expansion of UCEs during tetrapod evolution and the substitution rate in UCEs showed a significant decline in tetrapods compared with fugu, suggesting they were exapted in tetrapods. They considered it unlikely that these elements are ancient but evolved at a higher rate in teleost fishes. In this study, we investigated the evolution of UCEs in a cartilaginous fish, the elephant shark and show that nearly half the UCEs were present in the jawed vertebrate ancestor. The substitution rate in UCEs is higher in fugu than in elephant shark, and approximately one-third of ancient UCEs have diverged beyond recognition in teleost fishes. These data indicate that UCEs have evolved at a higher rate in teleost fishes, which may have implications for their vast diversity and evolutionary success.

Mammalian genomes contain thousands of highly conserved sequences that are under evolutionary selection. Surprisingly, a major portion of them are located in the noncoding regions of the genomes. Functional assays of such conserved noncoding sequences have indicated that many of them function as transcriptional regulatory elements (Shin et al. 2005; Woolfe et al. 2005; Pennacchio et al. 2006). Because the amount of noncoding sequences shows a broad correlation with the developmental complexity of eukaryotes, it has been hypothesized that noncoding sequences have played an important role in the evolution of complex developmental programs and phenotypic diversity of organisms (Taft et al. 2007). Therefore, investigations of the evolution of the conserved mammalian sequences in other vertebrate lineages should provide useful insights into the origin and the role of these elements in the evolution of phenotypic diversity of vertebrates. In a recent study, Stephen et al. (2008) identified 13,736 ultraconserved elements (UCEs) in the human genome that are identical over  $\geq 100$  bp in at least three out of five placental mammals (human, mouse, rat, dog, and cow) and investigated the evolution of these sequences in opossum, chicken, frog, and fugu genomes. They found that about 40% of these eutherian UCEs were present before the speciation of ray-finned fishes; 30% appeared first in the tetrapod ancestor; 18% arose in the amniote ancestor; and 10% evolved in the therian ancestor (fig. 1). In addition, they found that the substitution rate in the UCEs that first appeared in ray-finned fishes reduced significantly in the frog and chicken lineages compared with the fish lineage. It was therefore hypothesized that the UCEs were exapted to perform novel functions in tetrapods. The alternative hypothesis that these elements were present in the ancestral vertebrate and have evolved rapidly in teleost fishes was rejected because it is less parsimonious.

In this study, we investigated the presence of eutherian UCEs in a cartilaginous fish, the elephant shark (*Callorhynchus milii*), for which a 1.4 $\times$  coverage genome sequence

was recently generated (Venkatesh et al. 2007). Cartilaginous fishes are the oldest group of living jawed vertebrates that diverged from bony vertebrates (ray-finned fishes, coelacanths, lungfishes, and tetrapods)  $\sim 528$  MYA (Hedges and Kumar 2003). Surprisingly, our study showed that a majority of UCEs were in fact present in the common ancestor of jawed vertebrates and that the ancient UCEs have accumulated substitutions at a higher rate in teleost fishes, so much so that one-third of them have diverged beyond recognition.

To identify the orthologs of eutherian UCEs in the elephant shark, we aligned the human genome and five other vertebrate genomes (opossum, chicken, frog, fugu, and elephant shark) using MULTIZ (Blanchette et al. 2004). Altogether orthologs for 5,677 (41.3%) UCEs could be identified in elephant shark (table 1). Because the 1.4 $\times$  assembly of the elephant shark represents approximately 75% of the genome, we estimate that the whole genome of the elephant shark contains  $\sim 7,600$  UCEs (55%). Interestingly, although  $>90\%$  of the UCEs identified in the elephant shark are conserved in frog, chicken, and opossum,  $\sim 35\%$  (2,009) of them are absent in fugu. Almost a similar number of UCEs are also not identifiable in the stickleback (1,802) and medaka (2,076) genomes. Thus, about one-third of ancient UCEs have evolved at a significantly higher rate and diverged beyond recognition or are lost in the teleost fish lineage.

In the absence of data from a cartilaginous fish, Stephen et al. had reported that 39.3% of UCEs (5,404) are present in the ray-finned fish, and 30.3% of UCEs subsequently appeared in the tetrapod ancestor. However, with the identification of 41.3% of UCEs in the elephant shark, we found that only 12.6% (1,736) UCEs first appeared in ray-finned fishes, and 18.3% (2,508) first appeared in the tetrapod ancestor. The proportion of UCEs that first appeared in the amniote ancestor is lowered from 18.5% to 16.1% (fig. 1). Thus, the inclusion of the elephant shark in the analysis shows that a major proportion of the UCEs were already present in the jawed vertebrate ancestor, and a lower proportion of UCEs were recruited in the bony vertebrate and tetrapod ancestors. Nevertheless, this finding is consistent with the notion that nearly half of eutherian UCEs were recruited during the evolution of different lineages of tetrapods and that most UCEs have expanded in

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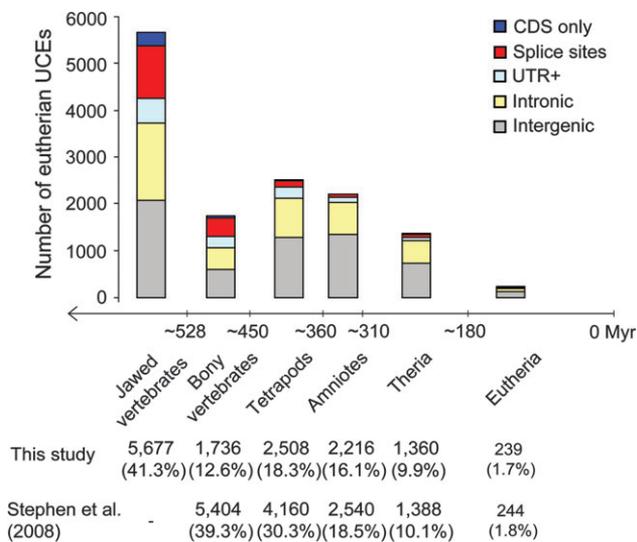


FIG. 1.—Distribution of UCEs reported by Stephen et al. (2008) and this study. No cartilaginous fish was analyzed in the former study, whereas the elephant shark was included in this study. The vertical bars show the genomic distribution of UCEs identified in this study. The estimated divergence times on the *x*-axis are based on molecular data (Hedges and Kumar 2003).

size. This study also shows that majority of the UCEs located within the protein-coding sequences are indeed ancient and were present in the common ancestor of jawed vertebrates, and confirms Stephen et al.'s finding that the new UCEs were added mainly in the noncoding (intergenic and intronic) sequences (fig. 1).

The substitution rate in UCEs in *fugu* is unusually higher compared with other vertebrates as shown by the extended branch length for *fugu* in a Neighbor-Joining tree built on ancient UCEs in all six species (fig. 2A and C). However, the higher evolutionary rate of UCEs in *fugu* seems to be the unique property of the UCEs and not a general property of the *fugu* genome, because the protein-coding sequences in *fugu* (CDSs; 271 genes, 198 kb aligned sequence) are evolving at a similar rate as in frog, chicken, and opossum (fig. 2B and C). Interestingly, the substitution rate (substitutions per site per  $10^9$  years) in the CDSs in the elephant shark (0.74) is lower than in all other vertebrates (*fugu*, 0.93; frog, 0.95; and chicken, 0.87; and opossum, 1.13). Previous studies of evolutionary rate of protein-coding sequences in cartilaginous fishes based on a few mitochondrial and nuclear genes had indicated that the rate of evolution of protein sequences in cartilaginous fishes is

**Table 1**  
**Eutherian UCEs (EU100+) Conserved in Vertebrates**

	Eutherian UCEs		Total Length (kb)	Average Identity (%)
	Number	%		
Human	13,736	100	2,131	NA
Opossum	13,184	96.0	2,035	94.7
Chicken	11,470	83.5	1,778	92.0
Frog	9,137	66.5	1,418	84.2
<i>Fugu</i>	5,404	39.3	773	74.1
Elephant shark <sup>a</sup>	5,677	41.3	837	79.8

<sup>a</sup> Elephant shark UCEs were identified in this study. The rest of the data are from Stephen et al. (2008).

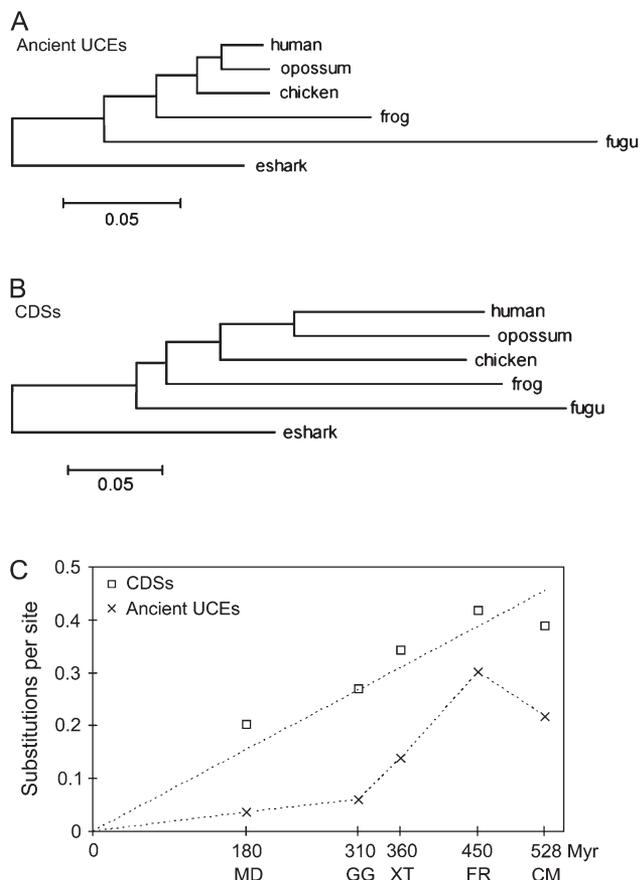


FIG. 2.—Substitution rates in UCEs and CDSs. The phylogenetic trees were generated using the Neighbor-Joining method and by specifying elephant shark as the outgroup. The standard errors for all branch lengths are less than  $10^{-3}$ . (A) Phylogenetic tree built from 3,297 ancient UCEs present in the elephant shark and other vertebrates (443 kb ungapped alignment). (B) Phylogenetic trees built from 271 elephant shark-coding sequences ranging in length from 228 bp to 2.1 kb and their orthologs in other species (198 kb ungapped alignment). eshark, elephant shark. Linearized versions of these trees show an incorrect placement of *fugu* as the most basal species (see supplementary fig. 1, Supplementary Material online). (C) Plot of divergence times (Hedges and Kumar 2003) versus substitution distance to human as derived from the trees in (A) and (B). MD, opossum; GG, chicken; XT, frog; FR, *fugu*; CM, elephant shark.

significantly lower than in mammals (Martin et al. 1992; Martin 1999; Kumazawa et al. 2000). Our analysis of a much larger set of protein sequences has confirmed that protein sequences in cartilaginous fishes have indeed been evolving at a slower rate than in other vertebrates.

The distribution of conservation across UCEs shows that the *fugu* UCEs are less conserved than the elephant shark UCEs across their entire length, with the level of divergence being relatively higher at the periphery (fig. 3). This indicates that the divergence of *fugu* UCEs has occurred mainly by accumulating a higher level of substitutions at the edges. It is likely that an acceleration of the decay at the edges might have led to the inability to recognize many ancient UCEs in *fugu*.

The investigation of the UCEs in the elephant shark has clearly shown that a substantial number of UCEs arose before the divergence of jawed vertebrates, and at least

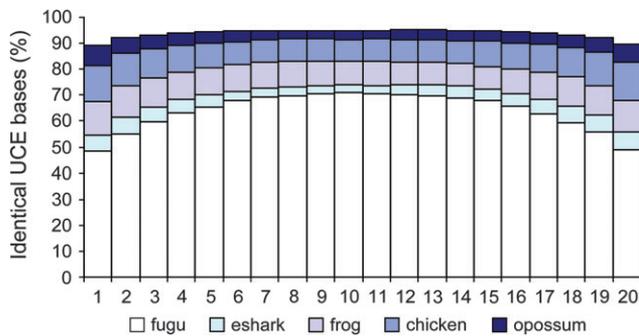


FIG. 3.—Distribution of conservation across UCEs. The eutherian UCEs conserved in each species (given in table 1) was divided into 20 equal sections and the percentage of identical bases was calculated for each section. *x*-axis, sections of UCEs; *y*-axis, percentage of identical bases.

one-third of ancient UCEs have diverged beyond recognition in teleost fishes. Furthermore, the ancient UCEs retained in teleost fishes have accumulated substitutions at an unusually higher rate compared with other vertebrates. Together, these data indicate that the UCEs have been evolving rapidly in the teleost fish lineage. The absence of a large number of ancient UCEs in teleost fishes is consistent with our previous observation that a significant number (~65%) of ancient conserved noncoding elements (>70% identical across >100 bp) present in the elephant shark and human have diverged beyond recognition in teleost fishes (Venkatesh et al. 2006). The rapid evolution of UCEs in teleost fishes could be the result of the whole-genome duplication in the ray-finned fish lineage (Christoffels et al. 2004; Jaillon et al. 2004). The genome duplication, which occurred in the teleost ancestor, might have relaxed the selective constraint on UCEs allowing them to diverge freely in teleosts. This hypothesis is supported by the observations that many duplicated genes in teleost fishes are evolving asymmetrically (Brunet et al. 2006; Steinke et al. 2006), and that the noncoding sequences in the duplicated HoxA clusters in zebrafish are less conserved compared with the noncoding sequences in the unduplicated HoxA cluster in the horn shark and human (Chiu et al. 2002). The consequences of the rapid divergence of ancient UCEs on the phenotypic evolution of teleost fishes are unclear. With about 27,000 living species, teleost fishes are the largest and most diverse group of vertebrates. The rapid evolution of UCEs might have influenced the vast divergence and speciation of teleost fishes.

## Methods

Pairwise alignments of human genome (hg18 assembly) with opossum (monDom4), chicken (galGal3), frog (xenTro2), fugu (fr2), stickleback (gasAcu1), and medaka (oryLat1) genomes were obtained from the UCSC Genome Browser (Karolchik et al. 2008). Pairwise alignment of human genome and elephant shark genome (version 1; Venkatesh et al. 2007) was generated using BlastZ (Schwartz et al. 2003). To identify UCEs present in elephant shark (“eshark”), multiple alignment of human

and five other vertebrate genomes was generated using MULTIZ/autoMZ (Blanchette et al. 2004) following the tree topology (((((human opossum) chicken) frog) fugu) eshark). Orthologous UCEs in elephant shark were identified using the same criteria as Stephen et al. (at least 20 alignable nucleotides). UCEs were annotated using the UCSC RefSeq gene track (dated August 20, 2008).

To build a phylogenetic tree that measures UCE substitution rates across six vertebrates, multiple alignments of the ancient UCEs were concatenated, and gapped columns were discarded. The multiple alignment was used to construct a Neighbor-Joining tree in MEGA4 (Tamura et al. 2007) with 1,000 bootstraps, Kimura 2-parameter model, and uniform rates among sites. To build a similar phylogenetic tree for coding sequences, protein sequences were first predicted from ~3,000 full-length elephant shark cDNA sequences using OrfPredictor (Min et al. 2005). These proteins were searched against the proteomes of human and other vertebrates by reciprocal BlastP ( $E < 10^{-20}$ ) and 271 proteins (unpublished; sequences available at <http://blast.fugu-sg.org/>, see Downloads) with orthologs in all six species were identified. Multiple alignments of the proteins from the six species were generated using ClustalW and converted to ungapped codon-based alignments by PAL2NAL (Suyama et al. 2006). Finally, the alignments were concatenated into a single alignment, and a Neighbor-Joining tree was constructed using similar settings as for UCEs.

## Supplementary Material

Supplementary figure 1 is available at *Molecular Biology and Evolution* (<http://www.mbe.oxfordjournals.org/>).

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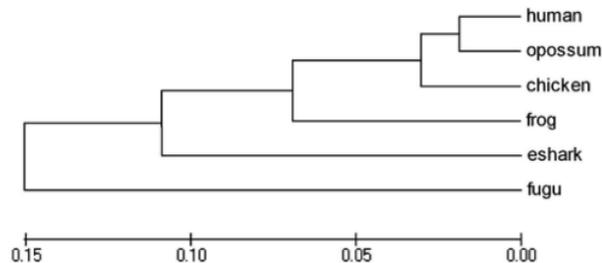
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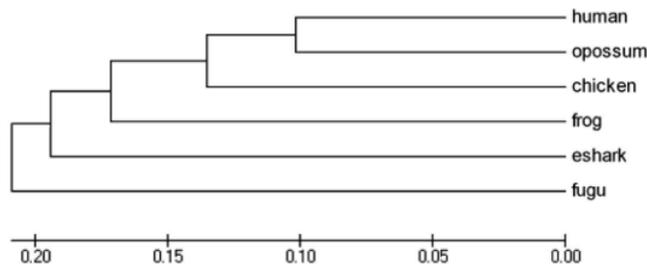
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## A Ancient UCEs



## B CDSs



Supplementary Fig. 1: Substitution rates in UCEs and CDSs. The phylogenetic trees were generated using Neighbor-Joining method and without specifying an outgroup. The distance in substitutions per site is shown below each tree. The standard errors for all branch lengths are less than  $10^{-3}$ . eshark, elephant shark. (A) Linearized phylogenetic tree built from 3,297 ancient UCEs present in the elephant shark and other vertebrates (443 kb ungapped alignment). (B) Linearized phylogenetic tree built from 271 elephant shark coding sequences ranging in length from 228 bp to 2.1 kb and their orthologs in other species (198 kb ungapped alignment). Note that because of the high substitution rates in UCEs in fugu, the phylogenetic tree for UCEs (A) is inconsistent with the species tree of these vertebrates; fugu is incorrectly placed as the most basal lineage. In the CDSs tree, largely because of the lower level of substitutions in the CDSs in the elephant shark, the elephant shark is incorrectly placed as a sister group of tetrapods (B).