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Genetic approaches to develop salt tolerant germplasm

Mark Tester*

King Abdullah University of Science and Technology, Thuwal, 23955-6900, Saudi Arabia

Abstract

Forty percent of the world's food is produced under irrigation, and this is directly threatened by over-exploitation and changes in the global environment. One way to address this threat is to develop systems for increasing our ability to use lower quality water, in particular saline water. Low cost partial desalination of brackish water, use of saline water for cooling and increases in the salinity tolerance of crops can all contribute to the development of this new agricultural system.

In this talk, the focus will be on the use of forward genetic approaches for discovery of genes related to salinity tolerance in barley and tomatoes. Rather than studying salinity tolerance as a trait in itself, we dissect salinity tolerance into a series of components that are hypothesised to contribute to overall salinity tolerance (following the paradigm of Munns & Tester, 2008).

For example, one significant component of tolerance of most crop plants to moderate soil salinity is due to the ability to maintain low concentrations of Na⁺ in the leaves, and much analysis of this aspect has been done (e.g. Roy *et al.*, 2013, 2014). A major site for the control of shoot Na⁺ accumulation is at the plasma membrane of the mature stele of the root. Alleles of *HKT*, a major gene underlying this transport process have been characterized and, in work led by Dr Rana Munns (CSIRO), have now been introgressed into commercial durum wheat and led to significantly increased yields in saline field conditions (Munns *et al.*, 2012).

The genotyping of mapping populations is now highly efficient. However, the ability to quantitatively phenotype these populations is now commonly limiting forward progress in plant science. The increasing power of digital imaging and computational technologies offers the opportunity to relieve this phenotyping bottleneck. The Plant Accelerator is a 4500m^2 growth facility that provides non-destructive phenotyping of large populations of plants (http://www.plantphenomics.org.au/). New genetic loci for previously under-studied components of salinity tolerance discovered using this new approach will be presented.

* Corresponding author. Tel.: +966 54 4700396 E-mail address:mark.tester@kaust.edu.sa The application of these technologies provides opportunities to significantly increase abiotic stress tolerance of crops, and thus contribute to increasing agricultural production in many regions, especially in the face of global environmental change. However, this needs to be tested in the field, such as done by Munns et al (2012) and Schilling et al. (2014). To this end, work will be described where mapping populations are grown in the field, and also grown in the Accelerator, and loci for traits are being compared with loci for tolerance in the field.

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References

Møller IS, Gilliham M, Jha D, Mayo GM, Roy SJ, Coates JC, Haseloff J, Tester M (2009) Plant Cell 21: 2163–2178; Munns R, Tester M (2008) Annu Rev Plant Biol 59: 651-681; Munns R, James RA, Xu B, Athman A, Jordans C, Conn SJ, Byrt CS, Hare RA, Tyerman SD, Tester M, Plett D, Gilliham M (2012) Nature Biotechnol 30: 360–364; Plett D, Safwat G, Shirley N, Møller IS, Gilliham M, Roy SJ, Jacobs A, Johnson A, Tester M (2010) PLoS ONE 5: e12571; Rajendran K, Tester M, Roy SJ (2009) Plant Cell Environ 32: 237-249; Roy SJ, Huang W, Wang X, Evrard A, Schmöckel S, Zafar ZU, Tester M (2013) Plant Cell Environ 35: 553-568; Roy S, Negrao, S, Tester M (2014) Curr Opinion Biotech 26: 115-124; Schilling RK, Marschner P, Shavrukov Y, Berger B, Tester M, Roy SJ, Plett DC (2014) Plant Biotech J 12: 378–386